

Supplementary Materials for

Human activities shape global patterns of decomposition rates in rivers

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Materials and Methods

Cellulose decomposition

We used a global dataset of cellulose-decomposition rates generated by a coordinated field experiment (Cellulose Decomposition Experiment [CELLDEX]) (*19*). Cotton strips were incubated in 514 flowing waters spanning 135 degrees of latitude by a consortium of over 150 peer-sourced researchers. Cotton strips are composed of cellulose, the primary constituent of most terrestrially derived leaf litter and the most abundant organic polymer on Earth; as such, cellulose is a plant polymer that is highly relevant for global biogeochemical cycles. The cottonstrip assay is an integrative measure of the activity of heterotrophic microbes and is highly sensitive to an array of environmental factors including nutrient concentrations, temperature, and pollutants (*24*). As used in our study the assay is not believed to be directly influenced by the feeding activity of macroinvertebrates. Cotton strips were deployed in 2015-2016 during periods of peak organic-matter inputs to flowing waters (e.g., autumn in temperate zones, dry season in tropical deciduous forests) at sites relatively free of major anthropogenic impacts. We typically chose stream orders 1-3 (*45*) and had sites located in each of Earth's major terrestrial biomes (*19*), and the cellulose-decomposition rate at each river was summarized as the exponential decay rate (*Kd*) of tensile-strength loss:

K_d =-ln(T_f/T_i)/*t*

where T_f is the final tensile strength of each cotton strip after incubation in the field, T_i is an average tensile-strength value of control strips not incubated in the field to establish initial tensile strength, and *t* the field incubation time in days (usually 21-30 days). The loss of tensile strength corresponds to the decomposition of the cotton fabric and is driven predominantly by the activity of microbes. Field and laboratory methods are detailed in (*19*, *24*).

Environmental data sources

For data on environmental variables other than *in-situ* water temperature, we relied on publicly available datasets with global coverage: 1) (*46*) for estimates of river yields of dissolved reactive phosphorus (kg DRP-P ha⁻¹ yr⁻¹) and nitrate+nitrite (kg NO_X-N ha⁻¹ yr⁻¹); 2) (47) for estimates of nitrogen (N) deposition; 3) (*48*) and (*49*) for estimates of phosphorus (P) deposition that we then interpolated; and 4) (*38*) for data on 96 variables summarized at the 12-digit hydrological scale or for the area upstream (HydroRIVERS: River ATLAS v10 lev12; HydroBASINS: BasinATLAS_v10_lev12) for either river reaches or corresponding subwatersheds, though all variables were not populated for all sub-watersheds. We excluded variables from HydroBASINS that were composite measures where we already included confounded variables (e.g., biome, human development index, and human footprint). We recorded temperature data with loggers for a subset (*n*=360) of the 514 rivers to determine the mean daily temperature of the river water during the cotton-strip incubation period.

Litter decomposition data

We used a global dataset of 3,216 unique estimates of litter-decomposition rates (as *K^d* using the equation above except that mass rather than tensile strength was used) for 125 plant genera and multiple experimental conditions (*27*) to independently validate whether our cellulose-decomposition model could explain rates of litter decomposition. These data are an

expanded version of the data published by LeRoy et al. (2020) (see data repository for complete data)(27). For each unique river reach sampled in the dataset, we averaged K_d estimates by each unique combination of leaf condition (i.e., leaves picked from the trees while still living or collected from the ground after senescence), plant genus, and direct feeding by detritivorous invertebrates (i.e., coarse-mesh which included invertebrates or fine-mesh litter bags which excluded invertebrates). We excluded any data for which we had 3 or fewer measurements of decomposition for a genus. The final dataset included 895 unique observations of 35 genera from 559 river reaches. All but 7 estimates of litter decomposition also included mean temperature during deployment, which we included as a predictor variable.

Leaf- and litter-trait data sources

We downloaded 384,252 records from 21,100 plant species and 4,557 genera of leaf traits related to nutrient, micronutrient, and structural compounds for leaves from the TRY plant-trait database (*31*). After filtering for traits describing the chemical constituency of plant leaves that we felt were most relevant for decomposition, the resulting database included average values for 7 traits representing 64 genera. Litter traits were assembled from 114 studies comprising 602 litter deployments of 172 genera in rivers (*43*). These trait values were joined by genus to the aforementioned empirical data on leaf litter. All genera for which we had litter-decomposition rates had data regarding either leaf or litter traits, and most included complete values for both. Details on filtering, aggregating, and variable selection as well as full datasets can be found in the data repository (*43*).

Data Analysis

Environmental data processing

At each river sampling location in the CELLDEX dataset, we combined temperature recorded during the experiment, extracted values from nutrient yield and deposition rasters, and attributes from HydroBASINS summarized by upstream watershed as well as the containing subwatershed. For HydroBASINS fields that were additionally available as monthly summaries (e.g., air temperature, potential evapotranspiration, snow coverage), we used both annual summaries and those from the month of deployment at each site as predictors in the BRT model. Variables from HydroBASINS were back-transformed into original units, and predictors with log-normal distributions were log₁₀ transformed. In total, we had 101 predictor variables for our cellulose-decomposition model.

Boosted-regression tree models

The choice between boosted regression tree (BRT) and other modeling techniques, such as Generalized Additive Models (GAMs) or neural networks depends on the specific characteristics of the data and the goals of the analysis. For our purposes, BRTs were an appropriate tool to answer our questions while addressing some of the complexity in our data. BRTs are recognized for their predictive accuracy, particularly in managing nonlinear relationships and interactions among predictors. The method is appropriate for handling missing data and outliers and processing large datasets. As the BRT constructs trees, it selects the most informative variables at each step, and assigns lower importance to the variables that contribute less to predictive performance. BRT is also resilient to irrelevant variables, as the boosting process assigns diminished weights to less informative variables and reduces their impact on the final model. BRTs can also capture interactions between variables and because the boosting process is adaptive, it allows the algorithm to focus on the most important variables and their interactions. Therefore, it reduces the risk of overfitting and improves the generalization of the model when challenged with new data. The learning rate of the BRT imposes a penalty on overfitting. Learning rates are often set to between 0.01 and 0.1, and ours was set to 0.001. Smaller learning rates put a penalty on the contribution of each tree, a technique that prevents the model from fitting the training data too closely.

In BRTs, "importance" refers to the degree of influence each predictor variable has on the predictive performance of the model and is normalized so the sum of all explanatory variable importance is 100. Variable importance in BRT is calculated based on how often a given variable is selected for: 1) splitting across all the trees, and 2) how much it contributes to the reduction in the model's loss function. Variables that are frequently chosen and contribute to improving model performance are considered more important. Higher importance values indicate features that have a greater impact on making accurate predictions. Detailed descriptions of the BRT approach are found in (*50*).

We used the gbm package in R (version 4.3.2) to build BRT models (*51, 52*) for cellulose decomposition and leaf-litter decomposition. Both BRT models were fitted with Gaussian distributions, learning rates of 0.001, and an interaction depth of 5. We initially used 20,000 trees in the cellulose model and the cross-validation determined the optional number of trees was 9,497. For the litter model, we initially ran 50,000 trees, and the optimal number was identified as 40,853. While BRT models handle variables with broad ranges, we ln-transformed K_d to facilitate the interpretation of results. The cellulose model used 101 explanatory variables (table s1) and the leaf-litter model used 17 explanatory variables (table s2). We assessed model explanatory power by calculating a pseudo- R^2 for each model and determined variable importance via permutation tests (*53*) (table 1). Explanatory variables with importance values greater than $1/n_{variable} * 100$ ($n_{variable} = total$ number of explanatory variables in the model) were included in trees more than would be expected from random chance and identified for further discussion (*54*). The importance threshold was 0.99 for the cellulose model and 5.88 for the leaflitter decomposition model. For the leaf litter model, two highly correlated explanatory variables (litter C:N and litter N content) fell just below the importance threshold but were discussed further because they each exceeded the threshold in other model runs and are well known to correlate with litter decomposition rates.

Output rasters of predicted cellulose-decomposition rates

Using the BRT models and data from the assembled spatial data layers, we predicted river K_d at the extent and at the resolution of the WorldClim rasters (global with 30 arc-second resolution; https://www.worldclim.org) using the raster package in R (*55*). In these output rasters, we did not predict K_d for sub-watersheds with ≤ 10 ha of sub-basin area, nor for Antarctica, which is not included in HydroATLAS. Importantly, we predicted *K^d* using a BRT model that included variables measured at each site in the original CELLDEX experiment (i.e., water temperatures and month of deployment), but those variables were not included in the generation of the global *K^d* map.

Validation of cellulose and leaf-litter BRT models

The spatial structure of the cellulose and leaf-litter datasets are quite different; therefore, we used different validation approaches for the cellulose and leaf-litter models. Because of the smaller dataset and hierarchical spatial structure of the cellulose-decomposition data (i.e., multiple streams measured by each partner), we performed a "leave-one-out" validation of the BRT by running 131 iterations of the model, each excluding one partner from the dataset. The goal was to assess the model's ability to predict the data of the omitted partner, measured through the calculation of root mean square error (RMSE). The average RMSE for the leave-one-out partner analysis was 1.08; in comparison, the BRT's cross-validation, which optimizes the number of trees directly in the code, yielded an RMSE of 0.93. The range of cellulose decomposition rates was 5.1 natural log units (K_d range 0.0012–0.20 d⁻¹). This analysis indicates that the model can predict cellulose decomposition rate with an accuracy of approximately $+/-1$ natural log unit and predictions in unsampled locations have similar accuracy to the model with all data included. For the larger, leaf-litter dataset compiled from published literature (n=895 decomposition rates), we randomly selected 80% of the data and used that to train the model, and we tested the model with the remaining 20% of the data. The average RMSE for the 80/20 analysis was 0.75 (n=20 random splits). In comparison, the BRT's cross-validation, which optimizes the number of trees directly in the code, yielded an RMSE of 0.76. The range of leaf litter decomposition rates was 5.9 natural log units (*K^d* range 0.005–0.18), which is much greater than the RMSE, indicating that the model is sufficiently accurate to make predictions of litter decomposition.

Data. All data and code for analyses and figures are available on GitHub (*43*).

Supplemental Acknowledgements: Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

Table S1.

Boosted-regression tree model importance values for cellulose decomposition rates (ln[*Kd*])), their description and the source of data. Importance values greater than 0.99 indicate that the variable was selected more than expected from random chance. The detailed information from the predictor variables derived from HydroBASINS can be found on their website. Variables that have similar names are typically referring to differences in the spatial or temporal characteristics of the variable. For example, air temperature tmp_dc_uyr is the annual average temperature for the total watershed upstream of sub-basin pour point, whereas tmp_dc_smx is the annual average temperature at the sub-basins pour point. If data were log transformed, "log", is written before the predictor variable text. The "Source" column denotes the origin of the data.

Table S2.

Boosted-regression tree model importance values for leaf-litter decomposition rates (ln[*Kd*]), their description and the source of data. Importance values greater than 5.88 indicate that the variable was selected more than expected from random chance. Additional information about plant traits can be found in the data repository (*43*) in the file "Litter_trait_review.csv" and more details about the TRY database are contained in (*31*).

Fig. S1.

Correlation plots of the relationship between the magnitude of predicted change in litterdecomposition rates in pine-dominated forests invaded by the pine bark beetle and watershed soil water content (A) and AET (B). Greater values indicate a higher magnitude increase in litter decomposition upon canopy replacement. Our forecasts predict insect-induced canopy replacement from pine to oak would approximately double mean decomposition rates (see main text). Though the relationships are highly variable, the associations between the predicted magnitude of change in decomposition and soil water and AET indicate drier subwatersheds are expected to have a larger change in decay rates than wetter sites.

References and Notes

- 1. J. Cebrian, Patterns in the fate of production in plant communities. *Am. Nat.* **154**, 449–468 (1999). [doi:10.1086/303244](http://dx.doi.org/10.1086/303244) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=10523491&dopt=Abstract)
- 2. T. J. Battin, R. Lauerwald, E. S. Bernhardt, E. Bertuzzo, L. G. Gener, R. O. Hall Jr., E. R. Hotchkiss, T. Maavara, T. M. Pavelsky, L. Ran, P. Raymond, J. A. Rosentreter, P. Regnier, River ecosystem metabolism and carbon biogeochemistry in a changing world. *Nature* **613**, 449–459 (2023). [doi:10.1038/s41586-022-05500-8](http://dx.doi.org/10.1038/s41586-022-05500-8) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=36653564&dopt=Abstract)
- 3. M. E. McClain, E. W. Boyer, C. L. Dent, S. E. Gergel, N. B. Grimm, P. M. Groffman, S. C. Hart, J. W. Harvey, C. A. Johnston, E. Mayorga, W. H. McDowell, G. Pinay, Biogeochemical hot spots and hot moments at the interface of terrestrial and aquatic ecosystems. *Ecosystems* **6**, 301–312 (2003). [doi:10.1007/s10021-003-0161-9](http://dx.doi.org/10.1007/s10021-003-0161-9)
- 4. E. R. Hotchkiss, R. O. Hall Jr., R. A. Sponseller, D. Butman, J. Klaminder, H. Laudon, M. Rosvall, J. Karlsson, Sources of and processes controlling $CO₂$ emissions change with the size of streams and rivers. *Nat. Geosci.* **8**, 696–699 (2015). [doi:10.1038/ngeo2507](http://dx.doi.org/10.1038/ngeo2507)
- 5. G. H. Allen, T. M. Pavelsky, Global extent of rivers and streams. *Science* **361**, 585–588 (2018). [doi:10.1126/science.aat0636](http://dx.doi.org/10.1126/science.aat0636) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=29954985&dopt=Abstract)
- 6. P. Regnier, P. Friedlingstein, P. Ciais, F. T. Mackenzie, N. Gruber, I. A. Janssens, G. G. Laruelle, R. Lauerwald, S. Luyssaert, A. J. Andersson, S. Arndt, C. Arnosti, A. V. Borges, A. W. Dale, A. Gallego-Sala, Y. Goddéris, N. Goossens, J. Hartmann, C. Heinze, T. Ilyina, F. Joos, D. E. Larowe, J. Leifeld, F. J. R. Meysman, G. Munhoven, P. A. Raymond, R. Spahni, P. Suntharalingam, M. Thullner, Anthropogenic perturbation of the carbon fluxes from land to ocean. *Nat. Geosci.* **6**, 597–607 (2013). [doi:10.1038/ngeo1830](http://dx.doi.org/10.1038/ngeo1830)
- 7. A. Marx, J. Dusek, J. Jankovec, M. Sanda, T. Vogel, R. van Geldern, J. Hartmann, J. A. C. Barth, A review of CO₂ and associated carbon dynamics in headwater streams: A global perspective. *Rev. Geophys.* **55**, 560–585 (2017). [doi:10.1002/2016RG000547](http://dx.doi.org/10.1002/2016RG000547)
- 8. P. A. Raymond, J. Hartmann, R. Lauerwald, S. Sobek, C. McDonald, M. Hoover, D. Butman, R. Striegl, E. Mayorga, C. Humborg, P. Kortelainen, H. Dürr, M. Meybeck, P. Ciais, P. Guth, Global carbon dioxide emissions from inland waters. *Nature* **503**, 355–359 (2013). [doi:10.1038/nature12760](http://dx.doi.org/10.1038/nature12760) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=24256802&dopt=Abstract)
- 9. V. Ferreira, A. C. Encalada, M. A. S. Graça, Effects of litter diversity on decomposition and biological colonization of submerged litter in temperate and tropical streams. *Freshw. Sci.* **31**, 945–962 (2012). [doi:10.1899/11-062.1](http://dx.doi.org/10.1899/11-062.1)
- 10. A. Bruder, M. H. Schindler, M. S. Moretti, M. O. Gessner, Litter decomposition in a temperate and a tropical stream: The effects of species mixing, litter quality and shredders. *Freshw. Biol.* **59**, 438–449 (2014). [doi:10.1111/fwb.12276](http://dx.doi.org/10.1111/fwb.12276)
- 11. J. C. Marks, Revisiting the fates of dead leaves that fall into streams. *Annu. Rev. Ecol. Evol. Syst.* **50**, 547–568 (2019). [doi:10.1146/annurev-ecolsys-110218-024755](http://dx.doi.org/10.1146/annurev-ecolsys-110218-024755)
- 12. M. A. S. Graça, The role of invertebrates on leaf litter decomposition in streams A review. *Int. Rev. Hydrobiol.* **86**, 383–393 (2001). [doi:10.1002/1522-](https://doi.org/10.1002/1522-2632(200107)86:4/5%3c383::AID-IROH383%3e3.0.CO;2-D) [2632\(200107\)86:4/5<383::AID-IROH383>3.0.CO;2-D](https://doi.org/10.1002/1522-2632(200107)86:4/5%3c383::AID-IROH383%3e3.0.CO;2-D)
- 13. T. V. Royer, G. W. Minshall, Controls on leaf processing in streams from spatial-scaling and hierarchical perspectives. *J. N. Am. Benthol. Soc.* **22**, 352–358 (2003). [doi:10.2307/1468266](http://dx.doi.org/10.2307/1468266)
- 14. J. J. Follstad Shah, J. S. Kominoski, M. Ardón, W. K. Dodds, M. O. Gessner, N. A. Griffiths, C. P. Hawkins, S. L. Johnson, A. Lecerf, C. J. LeRoy, D. W. P. Manning, A. D. Rosemond, R. L. Sinsabaugh, C. M. Swan, J. R. Webster, L. H. Zeglin, Global synthesis of the temperature sensitivity of leaf litter breakdown in streams and rivers. *Glob. Change Biol.* **23**, 3064–3075 (2017). [doi:10.1111/gcb.13609](http://dx.doi.org/10.1111/gcb.13609) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=28039909&dopt=Abstract)
- 15. M. Zhang, X. Cheng, Q. Geng, Z. Shi, Y. Luo, X. Xu, Leaf litter traits predominantly control litter decomposition in streams worldwide. *Glob. Ecol. Biogeogr.* **28**, 1469–1486 (2019). [doi:10.1111/geb.12966](http://dx.doi.org/10.1111/geb.12966)
- 16. L. Boyero, R. G. Pearson, M. O. Gessner, L. A. Barmuta, V. Ferreira, M. A. S. Graça, D. Dudgeon, A. J. Boulton, M. Callisto, E. Chauvet, J. E. Helson, A. Bruder, R. J. Albariño, C. M. Yule, M. Arunachalam, J. N. Davies, R. Figueroa, A. S. Flecker, A. Ramírez, R. G. Death, T. Iwata, J. M. Mathooko, C. Mathuriau, J. F. Gonçalves Jr., M. S. Moretti, T. Jinggut, S. Lamothe, C. M'Erimba, L. Ratnarajah, M. H. Schindler, J. Castela, L. M. Buria, A. Cornejo, V. D. Villanueva, D. C. West, A global experiment suggests climate warming will not accelerate litter decomposition in streams but might reduce carbon sequestration. *Ecol. Lett.* **14**, 289–294 (2011). [doi:10.1111/j.1461-0248.2010.01578.x](http://dx.doi.org/10.1111/j.1461-0248.2010.01578.x) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=21299824&dopt=Abstract)
- 17. G. Woodward, M. O. Gessner, P. S. Giller, V. Gulis, S. Hladyz, A. Lecerf, B. Malmqvist, B. G. McKie, S. D. Tiegs, H. Cariss, M. Dobson, A. Elosegi, V. Ferreira, M. A. S. Graça, T. Fleituch, J. O. Lacoursière, M. Nistorescu, J. Pozo, G. Risnoveanu, M. Schindler, A. Vadineanu, L. B.-M. Vought, E. Chauvet, Continental-scale effects of nutrient pollution on stream ecosystem functioning. *Science* **336**, 1438–1440 (2012). [doi:10.1126/science.1219534](http://dx.doi.org/10.1126/science.1219534) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=22700929&dopt=Abstract)
- 18. I. T. Handa, R. Aerts, F. Berendse, M. P. Berg, A. Bruder, O. Butenschoen, E. Chauvet, M. O. Gessner, J. Jabiol, M. Makkonen, B. G. McKie, B. Malmqvist, E. T. H. M. Peeters, S. Scheu, B. Schmid, J. van Ruijven, V. C. A. Vos, S. Hättenschwiler, Consequences of biodiversity loss for litter decomposition across biomes. *Nature* **509**, 218–221 (2014). [doi:10.1038/nature13247](http://dx.doi.org/10.1038/nature13247) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=24805346&dopt=Abstract)
- 19. S. D. Tiegs, D. M. Costello, M. W. Isken, G. Woodward, P. B. McIntyre, M. O. Gessner, E. Chauvet, N. A. Griffiths, A. S. Flecker, V. Acuña, R. Albariño, D. C. Allen, C. Alonso, P. Andino, C. Arango, J. Aroviita, M. V. M. Barbosa, L. A. Barmuta, C. V. Baxter, T. D. C. Bell, B. Bellinger, L. Boyero, L. E. Brown, A. Bruder, D. A. Bruesewitz, F. J. Burdon, M. Callisto, C. Canhoto, K. A. Capps, M. M. Castillo, J. Clapcott, F. Colas, C. Colón-Gaud, J. Cornut, V. Crespo-Pérez, W. F. Cross, J. M. Culp, M. Danger, O. Dangles, E. de Eyto, A. M. Derry, V. D. Villanueva, M. M. Douglas, A. Elosegi, A. C. Encalada, S. Entrekin, R. Espinosa, D. Ethaiya, V. Ferreira, C. Ferriol, K. M. Flanagan, T. Fleituch, J. J. Follstad Shah, A. Frainer Barbosa, N. Friberg, P. C. Frost, E. A. Garcia, L. García Lago, P. E. García Soto, S. Ghate, D. P. Giling, A. Gilmer, J. F. Gonçalves Jr, R. K. Gonzales, M. A. S. Graça, M. Grace, H.-P. Grossart, F. Guérold, V. Gulis, L. U. Hepp, S. Higgins, T. Hishi, J. Huddart, J. Hudson, S. Imberger, C. Iñiguez-Armijos, T. Iwata, D. J. Janetski, E. Jennings, A. E. Kirkwood, A. A. Koning, S. Kosten, K. A. Kuehn, H.

Laudon, P. R. Leavitt, A. L. Lemes da Silva, S. J. Leroux, C. J. LeRoy, P. J. Lisi, R. MacKenzie, A. M. Marcarelli, F. O. Masese, B. G. McKie, A. Oliveira Medeiros, K. Meissner, M. Miliša, S. Mishra, Y. Miyake, A. Moerke, S. Mombrikotb, R. Mooney, T. Moulton, T. Muotka, J. N. Negishi, V. Neres-Lima, M. L. Nieminen, J. Nimptsch, J. Ondruch, R. Paavola, I. Pardo, C. J. Patrick, E. T. H. M. Peeters, J. Pozo, C. Pringle, A. Prussian, E. Quenta, A. Quesada, B. Reid, J. S. Richardson, A. Rigosi, J. Rincón, G. Rîşnoveanu, C. T. Robinson, L. Rodríguez-Gallego, T. V. Royer, J. A. Rusak, A. C. Santamans, G. B. Selmeczy, G. Simiyu, A. Skuja, J. Smykla, K. R. Sridhar, R. Sponseller, A. Stoler, C. M. Swan, D. Szlag, F. Teixeira-de Mello, J. D. Tonkin, S. Uusheimo, A. M. Veach, S. Vilbaste, L. B. M. Vought, C.-P. Wang, J. R. Webster, P. B. Wilson, S. Woelfl, M. A. Xenopoulos, A. G. Yates, C. Yoshimura, C. M. Yule, Y. X. Zhang, J. A. Zwart, Global patterns and drivers of ecosystem functioning in rivers and riparian zones. *Sci. Adv.* **5**, eaav0486 (2019). [doi:10.1126/sciadv.aav0486](http://dx.doi.org/10.1126/sciadv.aav0486) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=30662951&dopt=Abstract)

- 20. D. M. Costello, S. D. Tiegs, L. Boyero, C. Canhoto, K. A. Capps, M. Danger, P. C. Frost, M. O. Gessner, N. A. Griffiths, H. M. Halvorson, K. A. Kuehn, A. M. Marcarelli, T. V. Royer, D. M. Mathie, R. J. Albariño, C. P. Arango, J. Aroviita, C. V. Baxter, B. J. Bellinger, A. Bruder, F. J. Burdon, M. Callisto, A. Camacho, F. Colas, J. Cornut, V. Crespo-Pérez, W. F. Cross, A. M. Derry, M. M. Douglas, A. Elosegi, E. de Eyto, V. Ferreira, C. Ferriol, T. Fleituch, J. J. Follstad Shah, A. Frainer, E. A. Garcia, L. García, P. E. García, D. P. Giling, R. K. Gonzales-Pomar, M. A. S. Graça, H.-P. Grossart, F. Guérold, L. U. Hepp, S. N. Higgins, T. Hishi, C. Iñiguez-Armijos, T. Iwata, A. E. Kirkwood, A. A. Koning, S. Kosten, H. Laudon, P. R. Leavitt, A. L. Lemes da Silva, S. J. Leroux, C. J. LeRoy, P. J. Lisi, F. O. Masese, P. B. McIntyre, B. G. McKie, A. O. Medeiros, M. Miliša, Y. Miyake, R. J. Mooney, T. Muotka, J. Nimptsch, R. Paavola, I. Pardo, I. Y. Parnikoza, C. J. Patrick, E. T. H. M. Peeters, J. Pozo, B. Reid, J. S. Richardson, J. Rincón, G. Risnoveanu, C. T. Robinson, A. C. Santamans, G. M. Simiyu, A. Skuja, J. Smykla, R. A. Sponseller, F. Teixeira-de Mello, S. Vilbaste, V. D. Villanueva, J. R. Webster, S. Woelfl, M. A. Xenopoulos, A. G. Yates, C. M. Yule, Y. Zhang, J. A. Zwart, Global patterns and controls of nutrient immobilization on decomposing cellulose in riverine ecosystems. *Global Biogeochemical Cycles* **36**, e2021GB007163 (2022). [doi:10.1126/sciadv.aav0486](http://dx.doi.org/10.1126/sciadv.aav0486)
- 21. C. M. Gough, Terrestrial primary production: Fuel for life. *Nature Education Knowledge* **3**, 28 (2011).
- 22. C. B. Field, M. J. Behrenfeld, J. T. Randerson, P. Falkowski, Primary production of the biosphere: Integrating terrestrial and oceanic components. *Science* **281**, 237–240 (1998). [doi:10.1126/science.281.5374.237](http://dx.doi.org/10.1126/science.281.5374.237) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=9657713&dopt=Abstract)
- 23. T. H. Huang, Y. H. Fu, P. Y. Pan, C. T. A. Chen, Fluvial carbon fluxes in tropical rivers. *Curr. Opin. Environ. Sustain.* **4**, 162–169 (2012). [doi:10.1016/j.cosust.2012.02.004](http://dx.doi.org/10.1016/j.cosust.2012.02.004)
- 24. S. D. Tiegs, J. E. Clapcott, N. A. Griffiths, A. J. Boulton, A standardized cotton-strip assay for measuring organic-matter decomposition in streams. *Ecol. Indic.* **32**, 131–139 (2013). [doi:10.1016/j.ecolind.2013.03.013](http://dx.doi.org/10.1016/j.ecolind.2013.03.013)
- 25. J. Mancuso, J. L. Tank, U. H. Mahl, A. Vincent, S. D. Tiegs, Monthly variation in organicmatter decomposition in agricultural stream and riparian ecosystems. *Aquat. Sci.* **85**, 83 (2023). [doi:10.1007/s00027-023-00975-7](http://dx.doi.org/10.1007/s00027-023-00975-7)
- 26. M. Ardón, L. H. Zeglin, R. M. Utz, S. D. Cooper, W. K. Dodds, R. J. Bixby, A. S. Burdett, J. Follstad Shah, N. A. Griffiths, T. K. Harms, S. L. Johnson, J. B. Jones, J. S. Kominoski, W. H. McDowell, A. D. Rosemond, M. T. Trentman, D. Van Horn, A. Ward, Experimental nitrogen and phosphorus enrichment stimulates multiple trophic levels of algal and detrital-based food webs: A global meta-analysis from streams and rivers. *Biol. Rev. Camb. Philos. Soc.* **96**, 692–715 (2020). [doi:10.1111/brv.12673](http://dx.doi.org/10.1111/brv.12673) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=33350055&dopt=Abstract)
- 27. C. J. LeRoy, A. L. Hipp, K. Lueders, J. J. Follstad Shah, J. S. Kominoski, M. Ardón, W. K. Dodds, M. O. Gessner, N. A. Griffiths, A. Lecerf, D. W. P. Manning, R. L. Sinsabaugh, J. R. Webster, Plant phylogenetic history explains in-stream decomposition at a global scale. *J. Ecol.* **108**, 17–35 (2020). [doi:10.1111/1365-2745.13262](http://dx.doi.org/10.1111/1365-2745.13262)
- 28. K. Yue, P. De Frenne, K. Van Meerbeek, V. Ferreira, D. A. Fornara, Q. Wu, X. Ni, Y. Peng, D. Wang, P. Heděnec, Y. Yang, F. Wu, J. Peñuelas, Litter quality and stream physicochemical properties drive global invertebrate effects on instream litter decomposition. *Biol. Rev. Camb. Philos. Soc.* **97**, 2023–2038 (2022). [doi:10.1111/brv.12880](http://dx.doi.org/10.1111/brv.12880) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=35811333&dopt=Abstract)
- 29. A. D. Rosemond, C. M. Pringle, A. Ramírez, M. J. Paul, J. L. Meyer, Landscape variation in phosphorus concentration and effects on detritus-based tropical streams. *Limnol. Oceanogr.* **47**, 278–289 (2002). [doi:10.4319/lo.2002.47.1.0278](http://dx.doi.org/10.4319/lo.2002.47.1.0278)
- 30. L. Boyero, M. A. S. Graça, A. M. Tonin, J. Pérez, A. J. Swafford, V. Ferreira, A. Landeira-Dabarca, M. A. Alexandrou, M. O. Gessner, B. G. McKie, R. J. Albariño, L. A. Barmuta, M. Callisto, J. Chará, E. Chauvet, C. Colón-Gaud, D. Dudgeon, A. C. Encalada, R. Figueroa, A. S. Flecker, T. Fleituch, A. Frainer, J. F. Gonçalves Jr., J. E. Helson, T. Iwata, J. Mathooko, C. M'Erimba, C. M. Pringle, A. Ramírez, C. M. Swan, C. M. Yule, R. G. Pearson, Riparian plant litter quality increases with latitude. *Sci. Rep.* **7**, 10562 (2017). [doi:10.1038/s41598-017-10640-3](http://dx.doi.org/10.1038/s41598-017-10640-3) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=28874830&dopt=Abstract)
- 31. J. Kattge, S. Díaz, S. Lavorel, I. C. Prentice, P. Leadley, G. Bönisch, E. Garnier, M. Westoby, P. B. Reich, I. J. Wright, J. H. C. Cornelissen, C. Violle, S. P. Harrison, P. M. Van Bodegom, M. Reichstein, B. J. Enquist, N. A. Soudzilovskaia, D. D. Ackerly, M. Anand, O. Atkin, M. Bahn, T. R. Baker, D. Baldocchi, R. Bekker, C. C. Blanco, B. Blonder, W. J. Bond, R. Bradstock, D. E. Bunker, F. Casanoves, J. Cavender-Bares, J. Q. Chambers, F. S. Chapin III, J. Chave, D. Coomes, W. K. Cornwell, J. M. Craine, B. H. Dobrin, L. Duarte, W. Durka, J. Elser, G. Esser, M. Estiarte, W. F. Fagan, J. Fang, F. Fernández-Méndez, A. Fidelis, B. Finegan, O. Flores, H. Ford, D. Frank, G. T. Freschet, N. M. Fyllas, R. V. Gallagher, W. A. Green, A. G. Gutierrez, T. Hickler, S. I. Higgins, J. G. Hodgson, A. Jalili, S. Jansen, C. A. Joly, A. J. Kerkhoff, D. Kirkup, K. Kitajima, M. Kleyer, S. Klotz, J. M. H. Knops, K. Kramer, I. Kühn, H. Kurokawa, D. Laughlin, T. D. Lee, M. Leishman, F. Lens, T. Lenz, S. L. Lewis, J. Lloyd, J. Llusià, F. Louault, S. Ma, M. D. Mahecha, P. Manning, T. Massad, B. E. Medlyn, J. Messier, A. T. Moles, S. C. Müller, K. Nadrowski, S. Naeem, Ü. Niinemets, S. Nöllert, A. Nüske, R. Ogaya, J. Oleksyn, V. G. Onipchenko, Y. Onoda, J. Ordoñez, G. Overbeck, W. A. Ozinga, S. Patiño, S. Paula, J. G. Pausas, J. Peñuelas, O. L. Phillips, V. Pillar, H. Poorter, L. Poorter, P. Poschlod, A. Prinzing, R. Proulx, A. Rammig, S. Reinsch, B. Reu, L. Sack, B. Salgado-Negret, J. Sardans, S. Shiodera, B. Shipley, A. Siefert, E. Sosinski, J. F. Soussana, E. Swaine, N. Swenson, K. Thompson, P. Thornton, M. Waldram, E. Weiher,

M. White, S. White, S. J. Wright, B. Yguel, S. Zaehle, A. E. Zanne, C. Wirth, TRY - A global database of plant traits. *Glob. Change Biol.* **17**, 2905–2935 (2011). [doi:10.1111/j.1365-2486.2011.02451.x](http://dx.doi.org/10.1111/j.1365-2486.2011.02451.x)

- 32. C. J. LeRoy, T. G. Whitham, S. C. Wooley, J. C. Marks, Within-species variation in foliar chemistry influences leaf-litter decomposition in a Utah river. *J. N. Am. Benthol. Soc.* **26**, 426–438 (2007). [doi:10.1899/06-113.1](http://dx.doi.org/10.1899/06-113.1)
- 33. A. Lecerf, E. Chauvet, Intraspecific variability in leaf traits strongly affects alder leaf decomposition in a stream. *Basic Appl. Ecol.* **9**, 598–605 (2008). [doi:10.1016/j.baae.2007.11.003](http://dx.doi.org/10.1016/j.baae.2007.11.003)
- 34. T. Sariyildiz, J. M. Anderson, Decomposition of sun and shade leaves from three deciduous tree species, as affected by their chemical composition. *Biol. Fertil. Soils* **37**, 137–146 (2003). [doi:10.1007/s00374-002-0569-y](http://dx.doi.org/10.1007/s00374-002-0569-y)
- 35. A. González-Hernández, R. Morales-Villafaña, M. E. Romero-Sánchez, B. Islas-Trejo, R. Pérez-Miranda, Modelling potential distribution of a pine bark beetle in Mexican temperate forests using forecast data and spatial analysis tools. *J. For. Res.* **31**, 649–659 (2018). [doi:10.1007/s11676-018-0858-4](http://dx.doi.org/10.1007/s11676-018-0858-4)
- 36. L. H. Fraser, H.A.L. Henry, C. N. Carlyle, S. R. White, C. Beierkuhnlein, J. F. Cahill, B. B. Casper, E. Cleland, S. L. Collins, J. S. Dukes, A. K. Knapp, E. Lind, R. Long, Y. Luo, P. B. Reich, M. D. Smith, M. Sternberg, R. Turkington, Coordinated distributed experiments: An emerging tool for testing global hypotheses in ecology and environmental science. *Front. Ecol. Environ.* **11**, 147–155 (2013). [doi:10.1890/110279](http://dx.doi.org/10.1890/110279)
- 37. B. Lehner, K. Verdin, A. Jarvis, New global hydrography derived from spaceborne elevation data. *Eos* **89**, 93–94 (2008). [doi:10.1029/2008EO100001](http://dx.doi.org/10.1029/2008EO100001)
- 38. B. Lehner, G. Grill, Global river hydrography and network routing: Baseline data and new approaches to study the world's large river systems. *Hydrol. Processes* **27**, 2171–2186 (2011). [doi:10.1002/hyp.9740](http://dx.doi.org/10.1002/hyp.9740)
- 39. M. O. Gessner, E. Chauvet, A case for using litter breakdown to assess functional stream integrity. *Ecol. Appl.* **12**, 498–510 (2002). [doi:10.1890/1051-](http://dx.doi.org/10.1890/1051-0761(2002)012%5b0498:ACFULB%5d2.0.CO;2) [0761\(2002\)012\[0498:ACFULB\]2.0.CO;2](http://dx.doi.org/10.1890/1051-0761(2002)012%5b0498:ACFULB%5d2.0.CO;2)
- 40. M. C. Jackson, O. L. F. Weyl, F. Altermatt, I. Durance, N. Friberg, A. J. Dumbrell, J. J. Piggott, S. D. Tiegs, K. Tockner, C. B. Krug, P. W. Leadley, G. Woodward, Recommendations for the next generation of global freshwater biological monitoring tools. *Adv. Ecol. Res.* **55**, 615–636 (2016). [doi:10.1016/bs.aecr.2016.08.008](http://dx.doi.org/10.1016/bs.aecr.2016.08.008)
- 41. K. A. Wilson, N. A. Auerbach, K. Sam, A. G. Magini, A. S. L. Moss, S. D. Langhans, S. Budiharta, D. Terzano, E. Meijaard, Conservation research is not happening where it is most needed. *PLOS Biol.* **14**, e1002413 (2016). [doi:10.1371/journal.pbio.1002413](http://dx.doi.org/10.1371/journal.pbio.1002413) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=27023288&dopt=Abstract)
- 42. A. D. Rosemond, J. P. Benstead, P. M. Bumpers, V. Gulis, J. S. Kominoski, D. W. P. Manning, K. Suberkropp, J. B. Wallace, Freshwater ecology. Experimental nutrient additions accelerate terrestrial carbon loss from stream ecosystems. *Science* **347**, 1142– 1145 (2015). [doi:10.1126/science.aaa1958](http://dx.doi.org/10.1126/science.aaa1958) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=25745171&dopt=Abstract)
- 43. D. Costello, J. P. Schmidt, C. Patrick, K. Capps, J. Follstad Shah, C. LeRoy, S. D. Tiegs, Data from: Human activities shape global patterns of decomposition rates in rivers, Zenodo (2024);<https://zenodo.org/records/11035638>
- 44. J. Cheng, B. Schloerke, B. Karambelkar, Y. Xie, leaflet: Create Interactive Web Maps with the JavaScript "Leaflet" Library, version 2.2.1, Comprehensive R Archive Network (2023); [https://cran.r-project.org/web/packages/leaflet/index.html.](https://cran.r-project.org/web/packages/leaflet/index.html)
- 45. A. N. Strahler, Quantitative analysis of watershed geomorphology. *Eos* **38**, 913–920 (1957).
- 46. R. W. McDowell, A. Noble, P. Pletnyakov, L. M. Mosley, Global database of diffuse riverine nitrogen and phosphorus loads and yields. *Geosci. Data J.* **8**, 132–143 (2021). [doi:10.1002/gdj3.111](http://dx.doi.org/10.1002/gdj3.111)
- 47. D. Ackerman, D. B. Millet, X. Chen, Global estimates of inorganic nitrogen deposition across four decades. *Global Biogeochem. Cycles* **33**, 100–107 (2019). [doi:10.1029/2018GB005990](http://dx.doi.org/10.1029/2018GB005990)
- 48. J. Brahney, N. Mahowald, D. S. Ward, A. P. Ballantyne, J. C. Neff, Is atmospheric phosphorus pollution altering global alpine Lake stoichiometry? *Global Biogeochem. Cycles* **29**, 1369–1383 (2015). [doi:10.1002/2015GB005137](http://dx.doi.org/10.1002/2015GB005137)
- 49. N. Mahowald, T. D. Jickells, A. R. Baker, P. Artaxo, C. R. Benitez-Nelson, G. Bergametti, T. C. Bond, Y. Chen, D. D. Cohen, B. Herut, N. Kubilay, R. Losno, C. Luo, W. Maenhaut, K. A. McGee, G. S. Okin, R. L. Siefert, S. Tsukuda, Global distribution of atmospheric phosphorus sources, concentrations and deposition rates, and anthropogenic impacts. *Global Biogeochem. Cycles* **22**, GB4026 (2008). [doi:10.1029/2008GB003240](http://dx.doi.org/10.1029/2008GB003240)
- 50. J. Elith, J. R. Leathwick, T. Hastie, A working guide to boosted regression trees. *J. Anim. Ecol.* **77**, 802–813 (2008). [doi:10.1111/j.1365-2656.2008.01390.x](http://dx.doi.org/10.1111/j.1365-2656.2008.01390.x) [Medline](http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&list_uids=18397250&dopt=Abstract)
- 51. R Core Team, R: A language and environment for statistical computing, version 4.3.2. Comprehensive R Archive Network (2023); [https://www.R-project.org/](https://www.r-project.org/)
- 52. G. Ridgeway, G. B. M. Developers, gbm: Generalized boosted regression models, version 2.1.9, Comprehensive R Archive Network (2024); [https://cran.r](https://cran.r-project.org/web/packages/gbm/index.html)[project.org/web/packages/gbm/index.html.](https://cran.r-project.org/web/packages/gbm/index.html)
- 53. L. Breiman, Random forests. *Mach. Learn.* **45**, 5–32 (2001). [doi:10.1023/A:1010933404324](http://dx.doi.org/10.1023/A:1010933404324)
- 54. A. M. Thorn, J. R. Thompson, J. S. Plisinski, Patterns and predictors of recent forest conversion in New England. *Land (Basel)* **5**, 30 (2016). [doi:10.3390/land5030030](http://dx.doi.org/10.3390/land5030030)
- 55. R. J. Hijmans, raster: Geographic data analysis and modeling, version 3.6-36, Comprehensive R Archive Network (2023); [https://cran.r](https://cran.r-project.org/web/packages/raster/index.html)[project.org/web/packages/raster/index.html.](https://cran.r-project.org/web/packages/raster/index.html)