

# UNIVERSIDAD REGIONAL AMAZÓNICA IKIAM

Facultad Ciencias de la Vida Ingeniería en Ecosistemas

# Effect of passive restoration on soil bacterial communities and factors associated in the major terrestrial biomes

Kassandra del Rocío Bazantes Saltos

16 de julio de 2021, ciudad de Tena, Napo, Ecuador



### Declaración de derecho de autor, autenticidad y responsabilidad

Tena, 16 de julio de 2021

Yo, Kassandra Del Rocío Bazantes Saltos con documento de identidad N° 1804711131, declaro que los resultados obtenidos en la investigación que presento en este documento final, previo a la obtención del título de Ingeniera en Ecosistemas, son absolutamente inéditos, originales, auténticos y personales.

En virtud de lo cual, el contenido, criterios, opiniones, resultados, análisis, interpretaciones, conclusiones, recomendaciones y todos los demás aspectos vertidos en la presente investigación son de mi autoría y de mi absoluta responsabilidad.

Por la favorable atención a la presente, suscribo de usted,

Atentamente,

Firma:

Kassandra Del Rocío Bazantes Saltos 1804711131



### Certificado de dirección de trabajo de integración curricular

Certifico que el trabajo de integración curricular titulado: Effect of passive restoration on soil bacterial communities and factors associated in the major terrestrial biomes, en la modalidad de: artículo de investigación, fue realizado por: Kassandra del Rocío Bazantes Saltos, bajo mi dirección. El mismo ha sido revisado en su totalidad y analizado por la herramienta de verificación de similitud de contenido; por lo tanto, cumple con los requisitos teóricos, científicos, técnicos, metodológicos y legales establecidos por la Universidad Regional Amazónica Ikiam, para su entrega y defensa.

Tena, 16 de julio de 2021



Roldán Torres Gutiérrez. Tutor

C.I: 0151345121



María Cristina Peñuela Mora. Co-tutora C.I: 1756861496

.....

#### www.ikiam.edu.ec

# CONTENIDO

1. Introduction
2. Materials and methods4
2.1. Searching literature and data extraction4
2.2. Meta-analysis6
3. Results7
3.1 Effect of passive restoration on soil microbial biomass, bacterial communities and soil properties recovery7
3.2. Factors affecting soil microbial biomass in a passive restoration
4. Discussion
4.1- Effect of passive restoration on soil microbial biomass, bacterial communities and soil properties recovery
4.2- Factors affecting soil microbial biomass in a passive restoration
5.Conclusion
6. References
7. Appendix

### Lista de figuras

### Abstract

Passive restauration is a common strategy for the recovery of the structure and functionality of forests throughout the world. The degree to which this has occurred is usually evaluated through the study of the plant and animal species present, and soil properties, while little attention has been paid to changes in the communities of soil microorganisms. In order to elucidate the influence of passive restoration in different terrestrial biomes on soil bacteria communities, microbial biomass, and the physical and chemical properties of soil, the present study undertook a meta-analysis of 82 articles to answer the following questions: (i) Does the resilience of soil microbiota and the physical and chemical properties of the soil after of a passive restoration process vary among terrestrial biomes? (ii) What are the levels of soil microbiota and the physical-chemical properties of restored forest compared with primary forest, within the different biomes? (iii) What are the most important soil properties contributing to the change on soil microbial biomass abundance in the passive restoration process? Our results showed that, in some biomes, the levels of soil properties, microbial biomass, and bacterial communities increased during the passive restoration process, but, even if the factors driving forest degradation are removed, the soil may not return to its original state. Moreover, we found that there are only moderate correlations between microbial C and N response ratios (RR) and SOC RR, in this restoration process.

**Keywords:** passive restoration, soil microbial biomass, soil bacterial community, soil physical-chemical properties.

vi

### Resumen

La restauración pasiva es una estrategia común para la recuperación de la estructura y funcionalidad de los bosques en todo el mundo. El grado en que esto ha ocurrido generalmente se evalúa mediante el estudio de las especies vegetales y animales presentes y las propiedades del suelo, mientras que se ha prestado poca atención a los cambios en las comunidades de microorganismos del suelo. Con el fin de dilucidar la influencia de la restauración pasiva sobre las comunidades de bacterias, la biomasa microbiana y las propiedades físicas y químicas del suelo, en diferentes biomas terrestres, el presente estudio realizó un metaanálisis de 82 artículos para responder a las siguientes preguntas: (i) ¿La resiliencia del microbiota del suelo y las propiedades físicas y químicas del suelo después de un proceso de restauración pasiva varían entre los biomas terrestres? (ii) ¿Cuáles son los niveles de microbiota del suelo y las propiedades físico-químicas del bosque restaurado en comparación con el bosque primario, dentro de los diferentes biomas? (iii) ¿Cuáles son las propiedades del suelo más importantes que contribuyen al cambio en la abundancia de la biomasa microbiana del suelo en el proceso de restauración pasiva? Los resultados mostraron que, en algunos biomas, los niveles de propiedades del suelo, biomasa microbiana y comunidades bacterianas aumentaron durante el proceso de restauración pasiva, pero, incluso si se eliminan los factores que impulsan la degradación forestal, es posible que el suelo no vuelva a su estado original. Además, se encontró que solo existen correlaciones moderadas entre los índices de respuesta (RR) de C y N microbianos y el RR de SOC, en este proceso de restauración.

**Palabras clave:** restauración pasiva, biomasa microbiana del suelo, comunidad bacteriana del suelo, propiedades físico-químicas del suelo.

vii

# Revista elegida

Nombre: Soil Biology and Biochemistry

Página web: Soil Biology and Biochemistry | Journal | ScienceDirect.com by Elsevier

# Effect of passive restoration on soil bacterial communities and factors associated in the major terrestrial biomes

Bazantes Kassandra <sup>a</sup>, Torres Roldán <sup>a</sup>, Peñuela María Cristina <sup>b</sup>

<sup>a</sup> Applied microbiology research group, Ikiam Amazon Regional University, Tena-Ecuador

<sup>b</sup> Tropical Ecosystems and Global Change Research group, Ikiam Amazon Regional University, Tena-Ecuador

### Abstract

Passive restauration is a common strategy for the recovery of the structure and functionality of forests throughout the world. The degree to which this has occurred is usually evaluated through the study of the plant and animal species present, and soil properties, while little attention has been paid to changes in the communities of soil microorganisms. In order to elucidate the influence of passive restoration in different terrestrial biomes on soil bacteria communities, microbial biomass, and the physical and chemical properties of soil, the present study undertook a meta-analysis of 82 articles to answer the following questions: (i) Does the resilience of soil microbiota and the physical and chemical properties of the soil after of a passive restoration process vary among terrestrial biomes? (ii) What are the levels of soil microbiota and the physical-chemical properties of restored forest compared with primary forest, within the different biomes? (iii) What are the most important soil properties contributing to the change on soil microbial biomass abundance in the passive restoration process? Our results showed that, in some biomes, the levels of soil properties, microbial biomass, and bacterial communities increased during the passive restoration process, but, even if the factors driving forest degradation are removed, the soil may not return to its original state. Moreover, we found that there are only moderate correlations between microbial C and N response ratios (RR) and SOC RR, in this restoration process.

### 1. Introduction

Land-use changes are the main causes of the loss of biodiversity and the structure of terrestrial ecosystems (Nepstad et al., 1999; Asner et al., 2009; Gibbs et al., 2010; Köhl et al., 2015). Restoration is an important strategy to recover the structure and functionality of ecosystems after disturbance (Benayas et al., 2009; Bullock et al., 2011) and passive restoration has been shown to be effective in the recovery of abandoned agricultural lands throughout the world (Cramer et al., 2008; Guariguata and Ostertag, 2011; Shimamoto et al., 2018), with benefits including maximizing biodiversity, provision of ecosystem services, landscape connectivity, and improving soil quality (Zhang, et al., 2011: Crouzeilles et al., 2015). Assessment of the effectiveness of passive restoration to return an ecosystem to its original state has been mainly based on the study of plants, animals, and soil properties (Liu, 2003; Long, 2014; Chazdon and Guariguata, 2016; Deng et al., 2017; Meli et al., 2017). Its ability to restore communities of soil microorganisms, however, has received little attention and is still uncertain.

Microbial biomass and community structure are vital in mediating biogeochemical cycles. Indeed, microbial biomass is the most active fraction of the soil organic matter. Bacteria are the major natural agents responsible for nitrogen fixation and transformation in forest ecosystems (Reed et al, 2011) and are considered to be highly important in decomposing dead fungal biomass and thus incorporate cellulose-derived organic matter into the soil (Štursová et al, 2012; Eichorst and Kuske, 2012; Brabcová et al. 2016; López-Mondéjar et al. 2016). Within the domain Bacteria, the phylums Proteobacteria (copiotrophic), Actinobacteria (copiotrophic) and Acidobacteria (oligotrophic) play a vital role in the carbon cycle and have a function in recovering soils as beneficial to soil nutrient cycling (Aislabie et al., 2013; Fierer et al., 2007; Huang et al., 2015; Kielak et al., 2016).

Therefore, understanding the changes in the soil microbial biomass and bacterial communities during passive restoration activity is essential to our comprehension of forests' responses to perturbations and restoration activities. It is currently understood that changes in soil microbial communities during secondary succession are influenced by several factors, such as pH, concentrations of carbon (C), nitrogen (N), and phosphorus (P)

(Fierer and Jackson, 2006; Banning et al., 2011), land-use history (Jangid et al, 2011), and plant-microbe interactions (Tarlera et al., 2008). These also vary across different spatial scales and ecosystems (Fierer and Jackson, 2006; Tripathi et al., 2016; Zeng et al, 2017; Cai et al., 2018).

Previous global analyses have examined changes in microbial communities and soil properties, according to types of disturbance, methods of restoration, types of ecosystems, and the state secondary succession (Zhao, et al., 2019, Zhou, 2020). For example, Zhao, et al., (2019) developed a global meta-analysis and found that soil microbial biomass, and soil bacterial and fungal abundance increased during the first 10 years, but decreased beyond 30 years. Zhou, et al., (2017) showed that the proportion of fungi to bacteria was significantly higher in forest than in grasslands and Zhou et al. (2018) found a significant correlation between microbial C:N ratio and soil pH and C: N. Nonetheless, these studies did not distinguish between primary forests and restored forests, which is key to understanding the stability and resilience of terrestrial ecosystems.

This meta-analysis aims to elucidate the influence of secondary succession on soil bacteria communities, microbial biomass, and the physical and chemical properties of the soil, in different terrestrial biomes. Thus, the following questions were addressed: (i) Does the resilience of soil microbiota and the physical and chemical properties of the soil after of a passive restauration process vary among terrestrial biomes? (ii) What are the levels of soil microbiota and the physical-chemical properties of restored forests compared with primary forest, within the different biomes? (iii) What are the most important soil properties contributing to the change on soil microbial biomass abundance in the passive restoration process? The goal is to contribute to understanding the responses of soil properties, microbial biomass, and soil bacterial communities to the passive restoration process, beyond the microhabitat scale, and the degree to which forest soils can recover after disturbance. This research will also provide information on soil bacterial communities' stability and resilience in major terrestrial biomes.

### 2. Materials and methods

### 2.1. Searching literature and data extraction

### 2.1.1. Literature search

We conducted an extensive literature survey through the ISI Web of Science and Google Scholar, using the following search term combinations: secondary succession, succession forest, restored and primary forest, natural succession, following agricultural abandonment, cropland, secondary forest regeneration, chrono sequence forest and soil microbial, microbial community, microbial biomass, soil bacterial and soil microorganisms.

We looked for the following variables: (a) soil properties, including soil pH, soil organic carbon (SOC), soil total nitrogen (TN) and soil carbon to nitrogen ratio (C:N ratio); (b) microbial biomass properties, namely microbial biomass carbon (MBC), microbial biomass nitrogen (MBN) and microbial C:N ratio (microbial C:N); (c) and six variables representing bacterial community compositions: relative abundances of Acidobacteria (AcidoB), Actinobacteria (ActinoB) and Proteobacteria (ProteoB), in the studies that met the following criteria:

### Selection criteria:

- 30-45-year-old forests whit passive restoration (restored forest) that were recovering from farmland or logging (degraded lands), with comparable data from forest in which no disturbance has ever been reported (primary forest), in the same abiotic and biotic conditions.
- Data of the chosen variables (means, observation numbers, and standard deviations or standard error), reported directly in the papers assessed.
- Data of the A horizon or a topsoil layer (0–10, or 0–15 cm), no others.
- Restored forests reported in the same article but with different environmental variables (e.g. passive restoration conducted under several geographical locations), were considered as independent studies.

### 2.1.2. Data extraction

- We mapped location of studies using QGIS 10.2.
- We chose the Terrestrial Biomes represented at least by three data points in each analysis variable (n > 3).
- We digitized figures with means and errors using PlotDigitizer 2.6.2 (http://plotdigitizer.sourceforge.net).
- We transformed standard errors (SEs) to SDs, when necessary, using the formula: SD = SE (n1/2).

We included 82 papers in this synthesis (Appendix), 39 grouped in "Degraded lands vs. Restored forest", and 43 in "Primary forest vs. Restored forest". These papers represent three terrestrial biomes: Temperate broadleaf & mixed forest, Tropical & subtropical moist broadleaf forest, and Montane grasslands & shrublands, adapted from Olson et al. (2001).



**Figure 1.** Spatial distribution of the present meta-analysis carried out on a global scale for the period 1997-2020.

### 2.2. Meta-analysis

#### 2.2.1 Response ratios

We used the response ratio (RR) (Gurevitch and Hedges, 1999; Hedges et al., 1999) to determine the effect of secondary succession (data: Degraded lands vs. Restored forest) and forest degradation (data: Primary forest vs. Restored forest) on which variable, and calculated by Eq. (1).

$$\ln RR = \ln \left(\frac{X_e}{X_c}\right) \tag{1}$$

Where Xe and Xc are the means of the concerned variable in the experimental group (restored forest) and control group (degraded lands or primary forest) respectively. We calculated the variance (v) associated with each InRR using the means, replicate numbers, and SDs of both experimental and control groups.

Because we considered subgroups (terrestrial biome types) in the meta-analysis and these subgroups were not randomly chosen, but represent fixed levels of a chosen characteristic to assess (Borenstein and Higgins 2013), we employed a fixed-effects-model and calculated the weighted mean of the natural logarithm of the response ratio (In RR++) (Eq. 2).

$$\ln RR_{++} = \frac{\sum_{i=1}^{k} w_i \, \ln RR}{\sum_{i=1}^{k} w_i}$$
(2)

where k is the number of observations and w is the equals the reciprocal of the variance (1/v).

Also, we transformed the InRR++ into the change percentage (A) to estimates the recovery percentage of the analyzed variables in the passive restoration process (Degraded land vs. Restored forest), and the percentage changes of the same variables concerning reference forest (Primary forest vs. Restored forest) (Eq. 3).

The recovery effect was considered significant when the confidence interval (CI) of the change percentage at the 95% level did not overlap with zero, (Koricheva et al., 2013).

$$A = \left(e^{\ln RR_{++}} - 1\right) \times 100\% \tag{3}$$

Therefore, in this meta-analysis, A+ represents a result that favors the experimental group (restored forest), while A- shows a result that favors the control group (degraded lands or primary forest).

### 2.2.2. Subgroup analysis and publication bias

We identified whether microbial biomass, bacterial community composition and soil properties change percentages differed among the subgroup (terrestrial biomes) by using a one-way analysis of variance (ANOVA or MetaAnova). Then, we estimated a linear regression analysis to examine the relationships between the RRs of microbial biomass and the RRs of soil properties for the two data: "Degraded lands vs. Restored forest" and "Primary forest vs. Restored forest". We used the Egger test to check for publication bias (p 0.01).

Meta-analysis was conducted with Rstudio software.

### 3. Results

# **3.1.** Effect of passive restoration on soil microbial biomass, bacterial communities and soil properties recovery

### Degraded lands vs. Restored forest

Microbial biomass, bacterial diversity, and soil properties levels were greater in restored forest than in degraded lands (Figure 2). Subgroup analysis and ANOVA revealed that microbial biomass carbon increments, as an effect of passive restoration, were different among the three biomes (p<0.05), in the following descending order: temperate broadleaf & mixed forest (29%); tropical & subtropical moist broadleaf forest (11%); montane grasslands & shrublands (4%). Microbial biomass nitrogen, microbial C:N ratio, bacterial communities and soil properties were only analyzed for temperate broadleaf and mixed forest, due to the lack of published data for other biomes.

Within bacterial phyla, forest restoration consistently increased the relative abundances of Acidobacteria, Actinobacteria, and Proteobacteria in the temperate broadleaf and mixed forest (<40%) (Figure 2). Moreover, restored forest of temperate broadleaf and mixed biome had 70% more SOC, 70% more TN, 60% greater C: N, and 37% higher pH than

comparable degraded lands. Similarly, the passive restoration of montane grasslands and shrublands biome also increased soil TN amount (60%) (Figure 3).



+ Temperate broadleaf & mixed forest

🕂 Tropical & subtropical moist broadleaf forests

**Figure 2.** Effects of passive restoration on microbial composition (Microbial biomass C: microbial biomass carbon, Microbial biomass N: microbial biomass nitrogen and Microbial C: N: microbial C:N ratio), and bacterial community (AdcidoB: Acidobacteria, ActinoB: Actinobacteria, ProteoB: Proteobacteria) with respect to degraded lands (Degraded lands Vs. Restored forest) and reference forest (Primary forest Vs. Restored forest). The bars represent the 95% confidence intervals (CIs). The vertical dashed lines are the reference of a response ratio of zero and the numbers in parentheses are sample sizes.

### Primary forest vs. Restored forest

Microbial biomass carbon was higher in restored forest when compared to primary forest in temperate broadleaf and mixed forest biome (Figure 2). In contrast, microbial biomass nitrogen was 25% lower in restored montane grasslands and shrublands soils, while in tropical and subtropical forests the soil microbial biomass differences between primary forest and restored forest were not significantly different (p>0.05) (Figure 2). On the other hand, Acidobacteria and Proteobacteria showed about 10% more in primary forest, compared to restored forest in temperate broadleaf and mixed forest, and tropical and subtropical forests. Conversely, Actinobacteria abundance depended on the biome (p<0.001), with 20% fewer in restored tropical and subtropical moist broadleaf forests, but 75% more in restored temperate forests, compared to examples of primary forest. Additionally, in the comparison of primary forest and restored forest, the metaANOVA showed that SOC and soil C: N levels were also influenced by biome type (p<0.05) (Figure 3). On the other hand, soil pH did not differ significantly between primary forest and restored forest (<3%).



**Figure 3.-** Effects of passive restoration on soil properties (Soil N: soil nitrogen; SOC: soil organic carbon; soil C: N: soil carbon to nitrogen ratio) with respect to degraded lands (Degraded lands Vs. Restored forest) and reference forest (Primary forest Vs. Restored forest). The bars represent the 95% confidence intervals (CIs). The vertical dashed lines are the reference of a response ratio of zero and the numbers in parentheses are sample sizes

### 3.2. Factors affecting soil microbial biomass in a passive restoration

There were only moderate correlations between microbial C and N response ratios (RR) and SOC RR, in the restored forest from degraded lands (Figure 4). In contrast, microbial C RR between primary forest and restored forest was highly correlated with SOC (R = 0.75, p = 0.03).



**Figure 4.-** Relationships between the response ratios (RRs) of soil properties (soil C: N: soil C:N ratio; SOC: soil organic carbon) and the RRs of microbial composition (microbial C: microbial biomass carbon; microbial N: microbial biomass nitrogen; microbial C: N: microbial C:N ratio). Gray color: Primary forest vs. Restored forest; Green color: Degraded lands vs. Restored forest.

### 4. Discussion

# 4.1. Effect of passive restoration on soil microbial biomass, bacterial communities and soil properties recovery.

We found that the microbial biomass, soil bacterial communities and soil properties were affected by the passive restoration process (Figure 2). Compared with degraded lands (farmland or logging), soils of restored forest were significantly richer, which was consistent with other meta-analyses conducted at the global scale (Fierer et al., 2009; Lange et al., 2015; Khlifa et al., 2017; Chen et al., 2019). Furthermore, we also found that increases in microbial biomass, soil bacterial communities and soil properties during regeneration depends on the geographic location of the restored soil. Thus, the trend for microbial biomass to return to natural levels also may depend on the biome analyzed (Figure 2). Our analysis showed that passive restoration successfully recovers the relative abundance of Acidobacteria, Actinobacteria, and Proteobacteria (Figure 2). Previous research suggested that there are competitive interactions between copiotrophs (Proteobacteria) and oligotrophs (Actinobacteria and Acidobacteria) during the restoration process (Ramirez et al., 2012; Guo et al., 2018; Chen et al, 2018). However, our results suggest that

Acidobacteria, Actinobacteria, and Proteobacteria showed a similar percentage of relative abundance recovery in restored forest. Moreover, this study showed that different biomes influence the magnitude of soil bacterial communities' recovery during regeneration.

### 4.1.1 Degraded lands vs. Restored forest

### Montane grassland and shrublands biome

Microbial biomass carbon recovery in restored forest of 4% compared to degraded lands was not significant in this biome, probably because the number of cases was low and therefore the 95% CI of MBC percentage of change was larger. However, this could be characteristic for this biome. Soil microorganisms of high-elevation ecosystems respond sensitively to changes in land use because low temperatures limit soil development, primary productivity, and nutrient cycling (Körner, 2003; Bühlmann et al., 2011). Moreover,

lower MBC recovery could be also associated with the forest revegetation direction, which can be affecting by restoration type (Zhang, et al., 2011; Cao, et al., 2017). Invasive species that are out of balance with the local ecology are a major problem in secondary forests and are usually not controlled in passive restoration (Anderson, 1995), with ecological impacts that range from local suppression of native species to whole-scale changes in the functioning of ecosystems (Mack, 1986; Chornesky and Randall, 2003). For example, the expansion of Green Alder (Alnus viridis) across the Alps is much faster than the re-growth of the primary montane forests there (Anthelme et al., 2007; Svensk, et al., 2021), leading to increases in total nitrogen (TN) given that alders fix nitrogen (Figure 3).

#### Temperate broadleaf and mixed forest

In the restored temperate forest, MBC and MBN level increased by 29% and 85% respectively, compared with degraded lands, which was significantly higher than other biomes (Figure 2). It is probably because the warm weather and humid climate are favorable for regeneration, which provides a positive rhizosphere effect on soil microorganisms (Rutigliano et al., 2004; Singh et al.,2004; Mackay et al., 2016). This enhancement of plant productivity and litter biomass would lead to accumulation of soil organic matter (Figure 3), which is an important substrate for soil microbes (Camenzind et al., 2018; Chen et al., 2018). Also, compared with montane grasslands, the leaf litter here decomposes faster and presents better nutrition (e.g., balanced nitrogen availability) (Kanerva and Smolander, 2007). In addition, our study demonstrated an increase in pH in response to passive restoration process, which is important because lead to changes in the microbial community (Fierer and Jackson, 2006; Rosenzweig et al., 2016).

The abundance of different bacteria phyla varied in the restored forest soil in this biome and there were differences between these and those of degraded lands. Proteobacteria are known to be copiotrophic, and their presence correlates positively with C and N pools (Goldfarb et al., 2011; Zhang et al., 2016), while both Actinobacteria and Acidobacteria are often considered oligotrophs and are adapted to resource-limited conditions (Fierer et al., 2007). Interestingly, there are not significant differences between the percentages of recovery of Acidobacteria and Proteobacteria, which may suggest that late-state restored

forests do not show a distinction between the recovery of copiotrophic groups and oligotrophic groups in this biome.

### Tropical and subtropical moist broadleaf forest

The present meta-analysis showed only small amounts of microbial biomass in soils of restored forests. Although the researchers found for this biome has focused mainly on MBC, this difference might be because soils in the (sub) tropical forests can be highly weathered and depleted in phosphorus, and constrain the accumulation of organic C, which can limit soil microbial growth (Nottingham et al., 2015; Vitousek et al., 2010; He et al., 2020; Miki et al, 2020). Thus, these characteristics prevent a positive correlation between high productivity and microbial biomass (Prach and Walker, 2020). This may also be related to rapid litter decomposition in this warm, moist environment, in which nutrients are assimilated almost immediately by plants (Palm et al., 2007). Although some studies report that plant diversity increases soil microbial biomass across a diverse range of terrestrial ecosystems (Zak et al., 2003; Lange et al. 2015; Chen et al., 2019), our study did not show this relation in the terrestrial biomes.

### 4.1.2. Primary forest vs. Restored forest

### Montane grassland and shrublands biome

MBN level of montane grasslands and shrublands was 25% lower in restored forest, compared primary forest, in contrast to the case of tropical and subtropical moist broadleaf forests. The difference between primary forest and restored forest in microbial biomass C: N levels was only 3% (Figure 2 and 3), suggesting that the passive restoration process does not change the proportion of bacterial to fungal biomass, but does change the capacity of microbial communities to decompose biomass, fix N, and mineralize N (Jia et al., 2005; Guo et al., 2018). Besides, forest regeneration affects the quality of SOM, preventing the availability of C, which was reflected in the SOC level for degraded lands (Figure 3). Although the difference in SOC between primary forest and restored forest is just 10%, it is the highest among the three biomes that were studied. This could be a consequence of

burning or grazing, which are factors that significantly influence the accumulation of dead biomass in primary grassland and shrublands (Baer, 2002). Despite this, our results showed the total SOC would be natural regeneration of this biome (Figure 3). This is a promising result since grasslands are important ecosystems for global C and N cycles, considering that they store 10–30% of SOC globally (Follett and Reed, 2010; Qiu et al., 2013).

### Temperate broadleaf and mixed forest

MBC level was significantly higher (185%) in restored forests than primary temperate forests. This may be due to the qualities and quantities of primary forest leaf litter, which contains more lignin and nitrogen (Figure 3), ultimately limiting microbial growth (Schipper et al., 2011; Griffiths and Philippot, 2013; Hawkes and Keitt, 2015; Zhang et al., 2016). Nonetheless, the main bacteria phyla relative abundance in the restored forest was similar that of primary forests. This may be due to the fine-textured soils of temperate forests that have silt contents between 50% and 80%, which is more favorable for bacterial growth because they improve the water-holding capacity and nutrient availability, in addition to protecting against bacterial grazers (Xu et al, 2018). Thus, for environmental restoration objectives, our results are encouraging since the major recycling pathways in temperate forests is microbial decomposition (Pausas and Bond, 2020).

### Tropical and subtropical moist broadleaf forest

Although there are only a few data to support a trend of MBC recovery back to levels found in primary forest, our results showed similar MBN, microbial C: N ratio, SOC and TN, Acidobacteria, Actinobacteria, and Proteobacteria abundance in the restored and primary tropical forests (Figure 3 and 4). Tropical/subtropical forest soil property recovery could be due to the structure of these forests. For instance, trees and other species that inhabit forest canopies (e.g., epiphytes, lianas), together with understory vegetation, may increase soil carbon and nutrient input quantity and quality during passive restoration process (Santiago and Wright, 2007). Furthermore, the recovery of the properties of these soils after a perturbation (e.g., farming) could be easier due to the lower natural levels of nutrients (Xu et al., 2018). The present study confirmed that passive restoration is a viable

way to mitigate anthropogenic impacts on tropical forest soils (Deng et al., 2016, Shimamoto et al., 2018), even though a full return to the enormous species richness of these forests will take a long time (Chazdon and Guariguata, 2016).

Our result showed that a passive restoration strategy does contribute to the recovery of the relative abundance of bacteria in the Acidobacteria and Proteobacteria to levels similar to those of primary forest. Actinobacteria, however, do not respond in same way (Figure 2). Curiously, a meta-analysis of bacterial responses to land-use changes across the tropics forest found a consistent decline in Acidobacteria and Proteobacteria and increases in Actinobacteria (Petersen et al., 2019).

### 4.2. Factors affecting soil microbial biomass in a passive restoration

We found that the response ratios of MBN and microbial C: N did not show correlations with the response ratios of SOC or soil C: N. The response ratio of SOC showed a positive correlation with the response ratio of MBC in degraded lands vs. restored forest, which means that increasing soil C inputs through plant root exudation and litter production may likely stimulate microbial biomass production.

### 5. Conclusion

Terrestrial biomes show different trends and magnitudes of recovery of microbial biomass, bacterial communities, and physical and chemical soil parameters. In some biomes, passive restoration may partially offset the losses of SOC, TN, soil C: N ratio, and bacterial communities, caused by land degradation. However, removing the human activity that caused the degradation may not necessarily cause the system to revert to its natural state, or recovery could be very slow (> 40 years). This last result may be verified by increasing the microbial diversity component in the present meta-analysis. Finally, new efforts are needed to quantify the effect of restoration on soil bacterial communities and its factors associated with all terrestrial biomes, including primary forests as experimental controls.

### 6. References

Aislabie, J., et al., 2013. Soil Microbes and Their Contribution to Soil Services. Ecosystem Services in New Zealand–Conditions and Trends. Manaaki Whenua Press, Lincoln, New Zealand, pp. 143–161.

Anderson, P., 1995. Ecological restoration and creation: a review. *Biological Journal* of the Linnean Society, 56(suppl\_1), 187-211.

Anthelme, F., Villaret, J., and Brun, J., 2007. Shrub encroachment in the Alps gives rise to the convergence of sub-alpine communities on a regional scale. J. Veg. Sci. 18, 355–362. doi: 10.1111/j.1654-1103.2007.tb02547.x

Asner, GP., Rudel, TK., Aide, TM., 2009. A contemporary assessment of change in humid tropical forests. Conserv Biol 23:1386–1395. (<u>https://doi.org/10.1111/j.1523-1739.2009.01333.x</u>).

Baer, S.G., Kitchen, D.J., Blair, J.M., Rice, C.W., 2002. Changes in ecosystem structure and function along a chronosequence of restored grasslands. Ecol. Appl. 12, 1688–1701.

Banning, N.C., Gleeson, D.B., Grigg, A.H., Grant, C.D., Andersen, G.L., Brodie, E.L., Murphy, D.V., 2011. Soil microbial community successional patterns during forest ecosystem restoration. Applied and Environmental Microbiology 77, 6158e6164.

Brabcová, V., Nováková, M., Davidová, A., Baldrian, P., 2016. Dead fungal mycelium in forest soil represents a decomposition hotspot and a habitat for a specific microbial community. *New Phytologist*, *210*(4), 1369-1381.

Benayas, J. M. R., Newton, A. C., Diaz, A., Bullock, J. M., 2009. Enhancement of biodiversity and ecosystem services by ecological restoration: a metaanalysis. *science*, *325*(5944), 1121-1124.

Bullock, J., Aronson, J., Newton, A., Pywell, R., Rey-Benayas, J., 2011. Restoration of ecosystem services and biodiversity: conflicts and opportunities. Trends in Ecology & Evolution, 26, 541–549. <u>https://doi.org/10.1016/j.tree.2011.06.011</u>

Cai, Z. Q., Zhang, Y. H., Yang, C., Wang, S., 2018. Land-use type strongly shapes community composition, but not always diversity of soil microbes in tropical China. *Catena*, *165*, 369-380.

Camenzind, T., Hättenschwiler, S., Treseder, K. K., Lehmann, A., Rillig, M. C., 2018. Nutrient limitation of soil microbial processes in tropical forests. Ecological Monographs, 88(1), 4–21. https://doi.org/10.1002/ ecm.1279

Cao, C., Zhang, Y., Cui, Z., Feng, S., Wang, T., Ren, Q., 2017. Soil bacterial community responses to revegetation of moving sand dune in semi-arid grassland. *Applied microbiology and biotechnology*, *101*(15), 6217-6228.

Chazdon, R. L., M. R. Guariguata., 2016. Natural regeneration as a tool for largescale forest restoration in the tropics: Prospects and challenges. Biotropica 48: 844–855

Chen, H., Li, D., Xiao, K., Wang, K., 2018. Soil microbial processes and resource limitation in karst and non-karst forests. Functional Ecology, 32(5), 1400–1409. <u>https://doi.org/10.1111/1365-2435.13069</u>

Chen, C., Chen, H. Y., Chen, X., Huang, Z., 2019. Meta-analysis shows positive effects of plant diversity on microbial biomass and respiration. *Nature communications*, *10*(1), 1-10.

Cramer, V. A., R. J. Hobbs, and R. J. STANDISH., 2008. What's new about old fields? Land abandonment and ecosystem assembly. Trends Ecol. Evol. 23: 104–112.

Chornesky, E. A., Randall, J. M., 2003. The threat of invasive alien species to biological diversity: setting a future course. *Annals of the Missouri Botanical Garden*, 67-76.

Deng, L., Liu, S.G., Kim, G.D., Sweeney, S., Peng, C.H., Shangguan, Z.P., 2017a. Past and future carbon sequestration benefits of China's grain for green program. Glob. Environ. Chang. 47, 13–20.

Eichorst, SA., Kuske, CR., 2012. Identification of cellulose-responsive bacterial and fungal communities in geographically and edaphically different soils by using stable isotope probing. Appl Environ Microbiol 78:2316 –2327. <u>https://doi.org/10.1128/AEM.07313-11</u>.

Fierer, N., Jackson RB., 2006. The diversity and biogeography of soil bacterial communities. Proc Natl Acad Sci U S A 103:626–631. https://doi.org/10.1073/pnas.0507535103.

Fierer, N., Bradford, M. A., Jackson, R. B., 2007. Toward an ecological classification of soil bacteria. *Ecology*, *88*(6), 1354-1364.

Fierer, N., Strickland, M.S., Liptzin, D., Bradford, M.A., Cleveland, C.C., 2009.Global patternsin belowground communities. Ecol. Lett. 12, 1238–1249

Follett, R.F., Reed, D.A., 2010. Soil carbon sequestration in grazing lands: societal benefits and policy implications. Rangeland Ecol. Manage. 63, 4–15.

Gibbs, HK., Ruesch, AS., Achard, F., Clayton, MK., Holmgren, P., Ramankutty, N., Foley, JA., 2010.Tropical forests were the primary sources of new agricultural land in the 1980s and 1990s. Proc Natl Acad Sci 107:16732–16737.

Goldfarb, K.C., Karaoz, U., Hanson, C.A., Santee, C.A., Bradford, M.A., Treseder, K.K., Wallenstein, M.D., Brodie, E.L., 2011. Differential growth responses of soil bacterial taxa to carbon substrates of varying chemical recalcitrance. Front. Microbiol. 2.

Griffiths, B.S., Philippot, L., 2013. Insights into the resistance and resilience of the soil microbial community. Fems Microbiol. Rev. 37 (2), 112–129.

Guariguata, M. R., R. Ostertag., 2001. Neotropical secondary forest succession: Changes in structural and functional characteristics. For. Ecol. Manage. 148: 185–206.

Guo, Y., Chen, X., Wu, Y., Zhang, L., Cheng, J., Wei, G., Lin, Y., 2018. Natural revegetation of a semiarid habitat alters taxonomic and functional diversity of soil microbial communities. *Science of the Total Environment*, *635*, 598-606.

Hawkes, C.V., Keitt, T.H., 2015. Resilience vs. historical contingency in microbial responses to environmental change. Ecol. Lett. 18 (7), 612–625.

He, X., Hou, E., Veen, G. F., Ellwood, M. F., Dijkstra, P., Sui, X., Chu, C., 2020. Soil microbial biomass increases along elevational gradients in the tropics and subtropics but not elsewhere. *Global Ecology and Biogeography*, *29*(2), 345-354.

Huang, X., Liu, L., Wen, T., Zhu, R., Zhang, J., and Cai, Z., 2015. Illumina MiSeq investigations on the changes of microbial community in the *Fusarium oxysporum* f.sp. cubense infected soil during and after reductive soil disinfestation. *Microbiol. Res.* 181, 33–42. doi: 10.1016/j.micres.2015.08.00

Jangid K, Williams M, Franzluebbers A, Schmidt T, Coleman D, Whitman W., 2011. Land-use history has a stronger impact on soil microbial community composition than aboveground vegetation and soil properties. Soil Biology and Biochemistry, 43(10), 2184-2193.

Jia, G. M., Cao, J. and Wang, G. 2005b. Influences of land management on soil nutrients and microbial biomass in the central Loess Plateau, Northwest China. Land Degrad. Dev. 16: 455–462

Kanerva, S., Smolander, A., 2007. Microbial activities in forest floor layers under silver birch, Norway spruce and Scots pine. *Soil Biology and Biochemistry*, *39*(7), 1459-1467.

Kielak, A. M., Barreto, C. C., Kowalchuk, G. A., van Veen, J. A., & Kuramae, E. E., 2016. The ecology of Acidobacteria: moving beyond genes and genomes. *Frontiers in microbiology*, *7*, 744.

Khlifa, R., Paquette, A., Messier, C., Reich, P. B. Munson, A. D., 2017. Do temperate tree species diversity and identity influence soil microbial community function and composition? Ecol. Evol. 7, 7965–7974.

Köhl, M., Lasco, R., Cifuentes, M., Jonsson, Ö., Korhonen, K. T., Mundhenk, P., ... & Stinson, G., 2015. Changes in forest production, biomass and carbon: Results from the 2015 UN FAO Global Forest Resource Assessment. *Forest Ecology and Management*, *352*, 21-34.

Ko"rner C., 2003. Alpine plant life. Berlin: Springer

Lange, M., Eisenhauer, N., Sierra, C. A., Bessler, H., Engels, C., Griffiths, R. I., ... Gleixner, G., 2015. Plant diversity increases soil microbial activity and soil carbon storage. *Nature communications*, 6(1), 1-8.

Liu, J., Ouyang, Z., Pimm, S.L., Raven, P.H., Wang, X., Miao, H., Han, N., 2003. Protecting China's biodiversity. Science 300, 1240–1241.

Long, H., 2014. Land use policy in China: introduction. Land Use Policy 40, 1–5

López-Mondéjar, R., Zühlke, D., Becher, D., Riedel, K., Baldrian, P., 2016. Cellulose and hemicellulose decomposition by forest soil bacteria proceeds by the action of structurally variable enzymatic systems. *Scientific reports*, 6(1), 1-12.

Mack, R. N. 1986. Alien plant invasion into the intermountain west: A case history. Pp. 191-213 in H. A. Mooney & J. A. Drake (editors), Ecology of Biological Invasions of North America and Hawaii. Springer-Verlag, New York

Mackay, J. E., Cunningham, S. C. & Cavagnaro, T. R., 2006. Riparian reforestation: are there changes in soil carbon and soil microbial communities? Sci. Total Environ.566–567, 960–967.

Meli, P., Holl, K. D., Rey Benayas, J. M., Jones, H. P., Jones, P. C., Montoya, D., Moreno Mateos, D. 2017. A global review of past land use, climate, and active vs. passive restoration effects on forest recovery. *Plos one*, *12*(2), e0171368.

Miki, T., Ushido, M., Fukui, S., Kondoh, M., 2010. Functional diversity of microbial decomposers facilitates plant coexistence in a plant-microbe-soil feedback model. Proc. Natl. Acad. Sci. U.S.A. 107, 14251–14256.

Nepstad, D, Verissimo, A., Alencar, A., Nobre, C., Lima, E., Lefebvre, P., Schlesinger, P., Potter, C., Moutinho, P., Mendoza, E., Cochrane, M., Brooks, V., 1999. Large-scale impoverishment of Amazonian forests by logging and fire. Nature 398, 505–508.

Nottingham, A. T., Whitaker, J., Turner, B. L., Salinas, N., Zimmermann, M., Malhi, Y., Meir, P., 2015. Climate warming and soil carbon in tropical forests: Insights from an elevation gradient in the Peruvian Andes. BioScience, 65(9), 906–921. <u>https://doi.org/10.1093/biosci/biv109</u>

Palm, C., Sanchez, P., Ahamed, S., Awiti, A., 2007. Soils: a contemporary perspective. Annu. Rev. Environ. Resour. 32, 99–129. <u>https://doi.org/10.1146/annurev.</u> <u>energy.31.020105.100307</u>

Pausas, J. G., Bond, W. J., 2020. On the three major recycling pathways in terrestrial ecosystems. *Trends in ecology & evolution*.

Petersen, I. A., Meyer, K. M., Bohannan, B. J., 2019. Meta-analysis reveals consistent bacterial responses to land use change across the tropics. *Frontiers in ecology and evolution*, *7*, 391.

Prach, K., Walker, L.R., 2011. Four opportunities for studies of ecological succession. Trends in Ecology and Evolution 26, 119e123.

Olson, D. M., Dinerstein, E., Wikramanayake, E. D., Burgess, N. D., Powell, G. V., Underwood, E. C., Kassem, K. R., 2001. Terrestrial Ecoregions of the World: A New Map of Life on Earth. A new global map of terrestrial ecoregions provides an innovative tool for conserving biodiversity. BioScience, 51(11), 933-938.

Qiu, L.,Wei,X.,Zhang,X., Cheng,J.,2013.Ecosystemcarbonandnitrogenaccumulation after grazing exclusion in semiarid grassland. PLoS One 8, e55433.

Ramirez, K.S., Craine, J.M., Fierer, N., 2012. Consistent effects of nitrogen amendments on soil microbial communities and processes across biomes. Glob. Chang. Biol. 18: 1918–1927. <u>https://doi.org/10.1111/j.1365-2486.2012.02639.x</u>

Reed, SC., Cleveland, CC., Townsend, AR., 2011. Functional ecology of free-living nitrogen fixation: a contemporary perspective. Annu Rev Ecol Evol Syst 42:489 –512.

Rosenzweig, S. T., Carson, M. A., Baer, S. G., Blair, J. M., 2016. Changes in soil properties, microbial biomass, and fluxes of C and N in soil following post-agricultural grassland restoration. *Applied soil ecology*, *100*, 186-194.

Rutigliano, F. A., D'ascoli, R., De Santo, A. V., 2004. Soil microbial metabolism and nutrient status in a Mediterranean area as affected by plant cover. *Soil Biology and Biochemistry*, *36*(11), 1719-1729.

Santiago, L. S., Wright, S. J., 2007. Leaf functional traits of tropical forest plants in relation to growth form. *Functional Ecology*, *21*(1), 19-27.

Schipper, L.A., Degens, B.P., Sparling, G.P., Duncan, L.C., 2001.Changes in microbial heterotrophic diversity along five plantsuccessional sequences. Soil Biol. Biochem. 33, 2093–2103

Svensk, M., Pittarello, M., Nota, G., Schneider, M. K., Allan, E., Mariotte, P., Probo, M., 2021. Spatial Distribution of Highland Cattle in Alnus viridis Encroached Subalpine Pastures.

Shimamoto, C. Y., Padial, A. A., da Rosa, C. M., Marques, M. C., 2018. Restoration of ecosystem services in tropical forests: a global meta-analysis. *PloS one*, *13*(12), e0208523.

Singh, B. K., Millard, P., Whiteley, A. S., Murrell, J. C., 2004. Unravelling rhizospheremicrobial interactions: opportunities and limitations. *Trends in microbiology*, *12*(8), 386-393. Štursová M, Žifc`áková L, Leigh MB, Burgess R, Baldrian P., 2012. Cellulose utilization in forest litter and soil: identification of bacterial and fungal decomposers. FEMS Microbiol Ecol 80:735–746. https://doi.org/ 10.1111/j.1574-6941.2012.01343.x.

Tarlera, S., Jangid, K., Ivester, A., Whitman, W., Williams, M., 2008. Microbial community succession and bacterial diversity in soils during 77,000 years of ecosystem development. FEMS Microbiology Ecology 64, 129e140.

Tripathi, R., Song, W., Slik, J., Sukri, R., Jaafar, S., Dong, K., Adams, J., 2016. Distinctive tropical forest variants have unique soil microbial communities, but not always low microbial diversity. Frontiers in Microbiology, 7, 376.

Uroz, S., Oger, P., Lepleux, C., Collignon, C., Frey-Klett, P., Turpault, MP., 2011. Bacterial weathering and its contribution to nutrient cycling in temperate forest ecosystems. Res Microbiol 162:820 – 831. https://doi.org/ 10.1016/j.resmic.2011.01.013.

Vitousek, P. M., Porder, S., Houlton, B. Z., Chadwick, O. A., 2010. Terrestrial phosphorus limitation: Mechanisms, implications, and ni- trogen–phosphorus interactions. Ecological Applications, 20(1), 5–15. <u>https://doi.org/10.1890/08-0127.1</u>

Xu, Z., Yu, G., Zhang, X., He, N., Wang, Q., Wang, S., ... Zhao, N., 2018. Biogeographical patterns of soil microbial community as influenced by soil characteristics and climate across Chinese forest biomes. *Applied Soil Ecology*, *124*, 298-305.

Zak, D. R., Holmes, W. E., White, D. C., Peacock, A. D., Tilman, D. 2003. Plant diversity, soil microbial communities, and ecosystem function: are there any links?. *Ecology*, *84*(8), 2042-2050.

Zhao, F. Z., Ren, C. J., Han, X. H., Yang, G. H., Wang, J., Doughty, R., Wang, Z. T., 2019. Trends in soil microbial communities in afforestation ecosystem modulated by aggradation phase. *Forest Ecology and Management*, *441*, 167-175.

Zhang, C., Xue, S., Liu, G. B., & Song, Z. L., 2011. A comparison of soil qualities of different revegetation types in the Loess Plateau, China. *Plant and Soil*, *347*(1), 163-178.

Zhang, C., Liu, G., Xue, S., Wang, G., 2016. Soil bacterial community dynamics reflectchanges in plant community and soil properties during the secondary succession ofabandoned farmland in the Loess Plateau. Soil Biol. Biochem. 97, 40–49.

Zeng, Q., An, S., Liu, Y., 2017. Soil bacterial community response to vegetation succession after fencing in the grassland of China. Sci. Total Environ. 609, 2–10.

Zhou, Z., Wang, C., Jiang, L., Luo, Y., 2017. Trends in soil microbial communities during secondary succession. *Soil Biology and Biochemistry*, *115*, 92-99.

Zhou, Z., Wang, C., Luo, Y., 2018. Effects of forest degradation on microbial communities and soil carbon cycling: a global meta-analysis. *Global Ecology and Biogeography*, 27(1), 110-124.

Zhou, Z., Wang, C., & Luo, Y., 2020. Meta-analysis of the impacts of global change factors on soil microbial diversity and functionality. *Nature communications*, *11*(1), 1-10.

## 7. Appendix

### Supplementary data

- 1. Allison, V. J., Miller, R. M., Jastrow, J. D., Matamala, R., Zak, D. R. 2005. Changes in soil microbial community structure in a tallgrass prairie chronosequence. *Soil Science Society of America Journal*, *69*(5), 1412-1421.
- Arunachalam, A., Pandey, H. N. 2003. Ecosystem restoration of Jhum fallows in Northeast India: microbial C and N along altitudinal and successional gradients. *Restoration Ecology*, 11(2), 168-173.
- Barber, N. A., Chantos-Davidson, K. M., Amel Peralta, R., Sherwood, J. P., Swingley, W. D. 2017. Soil microbial community composition in tallgrass prairie restorations converge with remnants across a 27-year chronosequence. *Environmental Microbiology*, 19(8), 3118-3131.
- 4. Barnett, S. E., Youngblut, N. D., Buckley, D. H. 2020. Soil characteristics and land-use drive bacterial community assembly patterns. *FEMS microbiology ecology*, *96*(1), fiz194.
- 5. Bogorodskaya, A. V., Shishikin, A. S. 2020. Dynamics, Structure, and Functional Activity of Microbial Biomass in Soils of Restoring Felled Areas in Fir Forests of the Yenisei Ridge. *Eurasian Soil Science*, *53*(1), 126-136.
- 6. Bossio, D. A., Girvan, M. S., Verchot, L., Bullimore, J., Borelli, T., Albrecht, A., ... Osborn, A. M. 2005. Soil microbial community response to land use change in an agricultural landscape of western Kenya. *Microbial ecology*, *49*(1), 50-62.
- 7. Cai, Z. Q., Zhang, Y. H., Yang, C., Wang, S. 2018. Land-use type strongly shapes community composition, but not always diversity of soil microbes in tropical China. *Catena*, *165*, 369-380.
- 8. Chabrerie, O., Laval, K., Puget, P., Desaire, S., Alard, D. 2003. Relationship between plant and soil microbial communities along a successional gradient in a chalk grassland in north-western France. *Applied Soil Ecology*, *24*(1), 43-56.
- Chai, Y., Cao, Y., Yue, M., Tian, T., Yin, Q., Dang, H., ... Wang, M. 2019. Soil abiotic properties and plant functional traits mediate associations between soil microbial and plant communities during a secondary forest succession on the Loess Plateau. *Frontiers in microbiology*, 10, 895.
- 10. Chauvat, M., Zaitsev, A. S., Wolters, V. 2003. Successional changes of Collembola and soil microbiota during forest rotation. *Oecologia*, *137*(2), 269-276
- 11. Chen, X., Su, Y., He, X., Wei, Y., Wei, W., & Wu, J. (2012). Soil bacterial community composition and diversity respond to cultivation in Karst ecosystems. *World Journal of Microbiology and Biotechnology*, *28*(1), 205-213.

- 12. Chen, F., Zheng, H., Zhang, K., Ouyang, Z., Li, H., Wu, B., & Shi, Q. (2013). Soil microbial community structure and function responses to successive planting of Eucalyptus. *Journal of Environmental Sciences*, *25*(10), 2102-2111.
- 13. Chen, W., Jiao, S., Li, Q., & Du, N. (2020). Dispersal limitation relative to environmental filtering governs the vertical small-scale assembly of soil microbiomes during restoration. *Journal of Applied Ecology*, *57*(2), 402-412.
- 14. Cui, Y., Fang, L., Guo, X., Wang, X., Wang, Y., Zhang, Y., & Zhang, X. (2019). Responses of soil bacterial communities, enzyme activities, and nutrients to agricultural-tonatural ecosystem conversion in the Loess Plateau, China. *Journal of Soils and Sediments*, 19(3), 1427-1440.
- da Silva, D. K. A., de Oliveira Freitas, N., de Souza, R. G., da Silva, F. S. B., de Araujo, A. S. F., & Maia, L. C. (2012). Soil microbial biomass and activity under natural and regenerated forests and conventional sugarcane plantations in Brazil. *Geoderma*, 189, 257-261.
- de Carvalho, T. S., Jesus, E. D. C., Barlow, J., Gardner, T. A., Soares, I. C., Tiedje, J. M., & Moreira, F. M. D. S. (2016). Land use intensification in the humid tropics increased both alpha and beta diversity of soil bacteria. *Ecology*, *97*(10), 2760-2771.
- Dhandapani, S., Ritz, K., Evers, S., Yule, C. M., & Sjögersten, S. (2019). Are secondary forests second-rate? Comparing peatland greenhouse gas emissions, chemical and microbial community properties between primary and secondary forests in Peninsular Malaysia. *Science of the Total Environment*, 655, 220-231.
- Dos Santos, U. J., De Medeiros, E. V., Duda, G. P., Marques, M. C., Souza, E. S. D., Brossard, M., & Hammecker, C. (2019). Land use changes the soil carbon stocks, microbial biomass and fatty acid methyl ester (FAME) in Brazilian semiarid area. Archives of Agronomy and Soil Science, 65(6), 755-769.
- 19. Elhottová, D., Szili-Kovács, T., & Tříska, J. (2002). Soil microbial community of abandoned sand fields. *Folia microbiologica*, *47*(4), 435-440.
- 20. Fu, Q., Liu, C., Ding, N., Lin, Y., Guo, B., Luo, J., & Wang, H. (2012). Soil microbial communities and enzyme activities in a reclaimed coastal soil chronosequence under rice–barley cropping. *Journal of Soils and Sediments*, *12*(7), 1134-1144.
- Garcia, C., Roldan, A., & Hernandez, T. (1997). Changes in microbial activity after abandonment of cultivation in a semiarid Mediterranean environment (Vol. 26, No. 1, pp. 285-292). American Society of Agronomy, Crop Science Society of America, and Soil Science Society of America
- 22. Gunina, A., Smith, A. R., Godbold, D. L., Jones, D. L., & Kuzyakov, Y. (2017). Response of soil microbial community to afforestation with pure and mixed species. *Plant and Soil*, *412*(1-2), 357-368.
- 23. Guo, Y., Hou, L., Zhang, Z., Zhang, J., Cheng, J., Wei, G., & Lin, Y. (2019). Soil microbial diversity during 30 years of grassland restoration on the Loess Plateau, China: Tight linkages with plant diversity. *Land Degradation & Development*, *30*(10), 1172-1182
- Guo-Mei, J. I. A., Zhang, P. D., Gang, W. A. N. G., Jing, C. A. O., Jing-Cheng, H. A. N., & Huang, Y. P. (2010). Relationship between microbial community and soil properties during natural succession of abandoned agricultural land. *Pedosphere*, 20(3), 352-360.
- 25. Guo, Y., Chen, X., Wu, Y., Zhang, L., Cheng, J., Wei, G., & Lin, Y. (2018). Natural revegetation of a semiarid habitat alters taxonomic and functional diversity of soil microbial communities. *Science of the Total Environment*, *635*, 598-606.

- 26. Guo, Z., Zhang, X., Green, S. M., Dungait, J. A., Wen, X., & Quine, T. A. (2019). Soil enzyme activity and stoichiometry along a gradient of vegetation restoration at the Karst Critical Zone Observatory in Southwest China. *Land Degradation & Development*, 30(16), 1916-1927.
- 27. He, R., Yang, K., Li, Z., Schädler, M., Yang, W., Wu, F., ... & Xu, Z. (2017). Effects of forest conversion on soil microbial communities depend on soil layer on the eastern Tibetan Plateau of China. *PloS one*, *12*(10), e0186053.
- 28. Jangid, K., Williams, M. A., Franzluebbers, A. J., Blair, J. M., Coleman, D. C., & Whitman, W. B. (2010). Development of soil microbial communities during tallgrass prairie restoration. *Soil Biology and Biochemistry*, *42*(2), 302-312.
- 29. Jangid, K., Williams, M. A., Franzluebbers, A. J., Schmidt, T. M., Coleman, D. C., & Whitman, W. B. (2011). Land-use history has a stronger impact on soil microbial community composition than aboveground vegetation and soil properties. *Soil Biology and Biochemistry*, *43*(10), 2184-2193.
- Jiang, J. P., Xiong, Y. C., Jiang, H. M., De-You, Y. E., Ya-Jie, S. O. N. G., & Feng-Min, L. I. (2009). Soil microbial activity during secondary vegetation succession in semiarid abandoned lands of Loess Plateau. *Pedosphere*, *19*(6), 735-747.
- 31. Jangid, K., Williams, M. A., Franzluebbers, A. J., Blair, J. M., Coleman, D. C., & Whitman, W. B. (2010). Development of soil microbial communities during tallgrass prairie restoration. *Soil Biology and Biochemistry*, *42*(2), 302-312.
- 32. Jiao, F., Wen, Z. M., & An, S. S. (2011). Changes in soil properties across a chronosequence of vegetation restoration on the Loess Plateau of China. *Catena*, *86*(2), 110-116.
- 33. Kasel, S., & Bennett, L. T. (2007). Land-use history, forest conversion, and soil organic carbon in pine plantations and native forests of south eastern Australia. *Geoderma*, *137*(3-4), 401-413.
- 34. Lan, G., Li, Y., Wu, Z., & Xie, G. (2017). Impact of tropical forest conversion on soil bacterial diversity in tropical region of China. *European Journal of Soil Biology*, *83*, 91-97.
- 35. Lauber, C. L., Ramirez, K. S., Aanderud, Z., Lennon, J., & Fierer, N. (2013). Temporal variability in soil microbial communities across land-use types. *The ISME journal*, 7(8), 1641-1650.
- Li, J., Liu, Y., Hai, X., Shangguan, Z., & Deng, L. (2019). Dynamics of soil microbial C: N: P stoichiometry and its driving mechanisms following natural vegetation restoration after farmland abandonment. *Science of the Total Environment*, 693, 133613.
- 37. Li, J., Shangguan, Z., & Deng, L. (2020). Dynamics of soil microbial metabolic activity during grassland succession after farmland abandonment. *Geoderma*, *363*, 114167.
- Li, W., Chen, X., Liu, M., Kuzyakov, Y., Jiang, C., Wu, M., & Li, Z. (2017). Shifts in microbial communities with increasing soil fertility across a chronosequence of paddy cultivation in subtropical China. *Applied Soil Ecology*, *120*, 153-159.
- 39. Liang, Y., Pan, F., He, X., Chen, X., & Su, Y. (2016). Effect of vegetation types on soil arbuscular mycorrhizal fungi and nitrogen-fixing bacterial communities in a karst region. *Environmental Science and Pollution Research*, *23*(18), 18482-18491.
- 40. Lie, X. I. A. O., LIU, G. B., Sha, X. U. E., & Zhang, C. (2013). Soil microbial community composition during natural recovery in the Loess Plateau, China. *Journal of Integrative Agriculture*, *12*(10), 1872-1883.

- Liu, G. Y., Chen, L. L., Shi, X. R., Yuan, Z. Y., Yuan, L. Y., Lock, T. R., & Kallenbach, R. L. (2019). Changes in rhizosphere bacterial and fungal community composition with vegetation restoration in planted forests. *Land Degradation & Development*, 30(10), 1147-1157.
- 42. Liu, Y., Zhu, G., Hai, X., Li, J., Shangguan, Z., Peng, C., & Deng, L. (2020). Long-term forest succession improves plant diversity and soil quality but not significantly increase soil microbial diversity: Evidence from the Loess Plateau. *Ecological Engineering*, *142*, 105631.
- 43. Liu, J., Jia, X., Yan, W., Zhong, Y., & Shangguan, Z. (2020). Changes in soil microbial community structure during long-term secondary succession. *Land Degradation & Development*, *31*(9), 1151-1166.
- 44. Liu, R., Zhou, X., Wang, J., Shao, J., Fu, Y., Liang, C., ... & Bai, S. H. (2019). Differential magnitude of rhizosphere effects on soil aggregation at three stages of subtropical secondary forest successions. *Plant and Soil*, 436(1), 365-380.
- 45. Maithani, K., Tripathi, R. S., Arunachalam, A., & Pandey, H. N. (1996). Seasonal dynamics of microbial biomass C, N and P during regrowth of a disturbed subtropical humid forest in north-east India. *Applied Soil Ecology*, 4(1), 31-37.
- 46. Mao, D. M., Min, Y. W., Yu, L. L., Martens, R., & Insam, H. (1992). Effect of afforestation on microbial biomass and activity in soils of tropical China. *Soil Biology and Biochemistry*, *24*(9), 865-872.
- 47. Moore-Kucera, J., & Dick, R. P. (2008). PLFA profiling of microbial community structure and seasonal shifts in soils of a Douglas-fir chronosequence. *Microbial ecology*, *55*(3), 500-511.
- Ovsepyan, L., Kurganova, I., de Gerenyu, V. L., & Kuzyakov, Y. (2019). Recovery of organic matter and microbial biomass after abandonment of degraded agricultural soils: the influence of climate. *Land Degradation & Development*, 30(15), 1861-1874.
- 49. Pan, F., Zhang, W., Liang, Y., Liu, S., & Wang, K. (2018). Increased associated effects of topography and litter and soil nutrients on soil enzyme activities and microbial biomass along vegetation successions in karst ecosystem, southwestern China. Environmental Science and Pollution Research, 25(17), 16979-16990.
- 50. Pressler, Y., Zhou, J., He, Z., Van Nostrand, J. D., & Smith, A. P. (2020). Postagricultural tropical forest regeneration shifts soil microbial functional potential for carbon and nutrient cycling. *Soil Biology and Biochemistry*, *145*, 107784.
- 51. Qi, L., & Yang, J. (2017). Microbial community composition regulates SOC decomposition response to forest conversion in a Chinese temperate forest. *Ecological research*, *32*(2), 163-172.
- 52. Ren, C., Chen, J., Deng, J., Zhao, F., Han, X., Yang, G., ... & Ren, G. (2017). Response of microbial diversity to C: N: P stoichiometry in fine root and microbial biomass following afforestation. *Biology and Fertility of Soils*, *53*(4), 457-468.
- 53. Ren, C., Wang, T., Xu, Y., Deng, J., Zhao, F., Yang, G., ... & Ren, G. (2018). Differential soil microbial community responses to the linkage of soil organic carbon fractions with respiration across land-use changes. *Forest Ecology and Management*, 409, 170-178.
- 54. Ren, C., Liu, W., Zhao, F., Zhong, Z., Deng, J., Han, X., ... & Ren, G. (2019). Soil bacterial and fungal diversity and compositions respond differently to forest development. *Catena*, *181*, 104071.

- 55. Rosenzweig, S. T., Carson, M. A., Baer, S. G., & Blair, J. M. (2016). Changes in soil properties, microbial biomass, and fluxes of C and N in soil following post-agricultural grassland restoration. *Applied soil ecology*, *100*, 186-194.
- 56. Saynes, V., Hidalgo, C., Etchevers, J. D., & Campo, J. E. (2005). Soil C and N dynamics in primary and secondary seasonally dry tropical forests in Mexico. *Applied Soil Ecology*, *29*(3), 282-289.
- Singh, K. D., Arifin, A., Radziah, O., Shamshuddin, J., Hazandy, A. H., Majid, N. M., ... & Keeren, S. R. (2013). Status of soil microbial population, enzymatic activity and biomass of selected natural, secondary and rehabilitated forests. *American Journal* of Environmental Sciences, 9(4), 301-309.
- Silva, C. F. D., Pereira, M. G., Miguel, D. L., Feitora, J. C. F., Loss, A., Menezes, C. E. G., & Silva, E. M. R. D. (2012). Total organic carbon, microbial biomass and soil enzyme activity areas of agriculture, forestry and grassland in the middle Valley of Paraíba do Sul River (RJ). *Revista Brasileira de Ciência do Solo*, *36*(6), 1680-1689.
- 59. Song, P., Ren, H., Jia, Q., Guo, J., Zhang, N., & Ma, K. (2015). Effects of historical logging on soil microbial communities in a subtropical forest in southern China. *Plant and soil*, *397*(1-2), 115-126.
- 60. Song, M., Peng, W., Du, H., & Xu, Q. (2019). Responses of soil and microbial C: N: P stoichiometry to vegetation succession in a Karst Region of Southwest China. *Forests*, *10*(9), 755.
- 61. Song, Z., Liu, G., & Zhang, C. (2019). Response of rhizosphere microbial communities to plant succession along a grassland chronosequence in a semiarid area. *Journal of Soils and Sediments*, 19(5), 2496-2508.
- Susyan, E. A., Wirth, S., Ananyeva, N. D., & Stolnikova, E. V. (2011). Forest succession on abandoned arable soils in European Russia–Impacts on microbial biomass, fungal-bacterial ratio, and basal CO2 respiration activity. *European Journal of Soil Biology*, 47(3), 169-174.
- Tripathi, B. M., Song, W., Slik, J. W. F., Sukri, R. S., Jaafar, S., Dong, K., & Adams, J. M. (2016). Distinctive tropical forest variants have unique soil microbial communities, but not always low microbial diversity. *Frontiers in microbiology*, *7*, 376.
- 64. van der Wal, A., van Veen, J. A., Smant, W., Boschker, H. T., Bloem, J., Kardol, P., ...
  & de Boer, W. (2006). Fungal biomass development in a chronosequence of land abandonment. *Soil Biology and Biochemistry*, *38*(1), 51-60.
- 65. Vasconcellos, R. D. F., Bonfim, J. A., Andreote, F. D., Mendes, L. W., Baretta, D., & Cardoso, E. J. B. N. (2013). Microbiological indicators of soil quality in a riparian forest recovery gradient. *Ecological engineering*, *53*, 313-320.
- 66. Wang, J., Liu, G., Zhang, C., Wang, G., Fang, L., & Cui, Y. (2019). Higher temporal turnover of soil fungi than bacteria during long-term secondary succession in a semiarid abandoned farmland. *Soil and Tillage Research*, 194, 104305.
- 67. Wang, K., Zhang, Y., Tang, Z., Shangguan, Z., Chang, F., Chen, Y., ... & Deng, L. (2019). Effects of grassland afforestation on structure and function of soil bacterial and fungal communities. *Science of the total environment*, *676*, 396-406.
- 68. Xue, L., Ren, H., Li, S., Leng, X., & Yao, X. (2017). Soil bacterial community structure and co-occurrence pattern during vegetation restoration in karst rocky desertification area. *Frontiers in microbiology*, *8*, 2377.

- 69. Yu, J., Liu, F., Tripathi, B. M., & Steinberger, Y. (2020). Changes in the composition of soil bacterial and fungal communities after revegetation with Caragana microphylla in a desertified semiarid grassland. *Journal of Arid Environments*, 182, 104262.
- 70. Zhang, H., Lu, J., & Zhao, S. (2009). Response of soil microorganisms to vegetational succession in Ziwuling Forest. *Frontiers of Environmental Science & Engineering in China*, 3(4), 421-427.
- 71. Zhang, C., Liu, G., Xue, S., & Zhang, C. (2012). Rhizosphere soil microbial properties on abandoned croplands in the Loess Plateau, China during vegetation succession. *European Journal of Soil Biology*, *50*, 127-136.
- 72. Zhang, Y., Cong, J., Lu, H., Yang, C., Yang, Y., Zhou, J., & Li, D. (2014). An integrated study to analyze soil microbial community structure and metabolic potential in two forest types. *PloS one*, *9*(4), e93773.
- 73. Zhang, C., Liu, G., Song, Z., Qu, D., Fang, L., & Deng, L. (2017). Natural succession on abandoned cropland effectively decreases the soil erodibility and improves the fungal diversity. *Ecological Applications*, *27*(7), 2142-2154.
- 74. Zhang, K., Cheng, X., Shu, X., Liu, Y., & Zhang, Q. (2018). Linking soil bacterial and fungal communities to vegetation succession following agricultural abandonment. *Plant and Soil*, 431(1), 19-36.
- 75. Zhang, W., Qiao, W., Gao, D., Dai, Y., Deng, J., Yang, G., ... & Ren, G. (2018). Relationship between soil nutrient properties and biological activities along a restoration chronosequence of Pinus tabulaeformis plantation forests in the Ziwuling Mountains, China. *Catena*, 161, 85-95.
- 76. Zhang, H., Xiong, X., Wu, J., Zhao, J., Zhao, M., Chu, G., ... & Zhang, D. (2019). Changes in Soil Microbial Biomass, Community Composition, and Enzyme Activities After Half-Century Forest Restoration in Degraded Tropical Lands. *Forests*, 10(12), 1124.
- 77. Zhang, K., Li, X., Cheng, X., Zhang, Z., & Zhang, Q. (2019). Changes in soil properties rather than functional gene abundance control carbon and nitrogen mineralization rates during long-term natural revegetation. *Plant and Soil*, 443(1), 293-306.
- 78. Zhao, J., Li, S., He, X., Liu, L., & Wang, K. (2014). The soil biota composition along a progressive succession of secondary vegetation in a karst area. *PloS one*, 9(11), e112436.
- 79. Zhao, F. Z., Ren, C. J., Han, X. H., Yang, G. H., Wang, J., & Doughty, R. (2018). Changes of soil microbial and enzyme activities are linked to soil C, N and P stoichiometry in afforested ecosystems. *Forest ecology and management*, *427*, 289-295.
- 80. Zhao, F. Z., Bai, L., Wang, J. Y., Deng, J., Ren, C. J., Han, X. H., ... & Wang, J. (2019). Change in soil bacterial community during secondary succession depend on plant and soil characteristics. *Catena*, *173*, 246-252.
- Zheng, H., Ouyang, Z. Y., Wang, X. K., Fang, Z. G., Zhao, T. Q., & Miao, H. (2005). Effects of regenerating forest cover on soil microbial communities: a case study in hilly red soil region, Southern China. *Forest Ecology and Management*, 217(2-3), 244-254.
- 82. Zhu, B., Li, Z., Li, P., Liu, G., & Xue, S. (2010). Soil erodibility, microbial biomass, and physical-chemical property changes during long-term natural vegetation restoration: a case study in the Loess Plateau, China. *Ecological Research*, 25(3), 531-541.

- 83. Zhu, H., He, X., Wang, K., Su, Y., & Wu, J. (2012). Interactions of vegetation succession, soil bio-chemical properties and microbial communities in a Karst ecosystem. *European Journal of Soil Biology*, *51*, 1-7.
- 84. Zieher, X. M. L., Vivanco, L., & Yahdjian, L. (2020). Soil bacterial communities remain altered after 30 years of agriculture abandonment in Pampa grasslands. *Oecologia*, 193(4), 959-968.